Dynamic nuclear polarization of spherical nanoparticles†

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Spherical silica nanoparticles of various particle sizes (~10 to 100 nm), produced by a modified Stoeber method employing amino acids as catalysts, are investigated using Dynamic Nuclear Polarization (DNP) enhanced Nuclear Magnetic Resonance (NMR) spectroscopy. This study includes ultra-sensitive detection of surface-bound amino acids and their supramolecular organization in trace amounts, exploiting the increase in NMR sensitivity of up to three orders of magnitude via DNP. Moreover, the nature of the silicon nuclei on the surface and the bulk silicon nuclei in the core (sub-surface) is characterized at atomic resolution. Thereby, we obtain unique insights into the surface chemistry of these nanoparticles, which might result in improving their rational design as required for promising applications, e.g. as catalysts or imaging contrast agents. The non-covalent binding of amino acids to surfaces was determined which shows that the amino acids not just function as catalysts but become incorporated into the nanoparticles during the formation process. As a result only three distinct Q-types of silica signals were observed from surface and core regions. We observed dramatic changes of DNP enhancements as a function of particle size, and very small particles (which suit in vivo applications better) were hyperpolarized with the best efficiency. Nearly one order of magnitude larger DNP enhancement was observed for nanoparticles with 13 nm size compared to particles with 100 nm size. We determined an approximate DNP penetration-depth (~4.2 or ~5.7 nm) for the polarization transfer from electrons to the nuclei of the spherical nanoparticles. Faster DNP polarization buildup was observed for larger nanoparticles. Efficient hyperpolarization of such nanoparticles, as achieved in this work, can be utilized in applications such as magnetic resonance imaging (MRI).

A Introduction

DNP increases the sensitivity of NMR experiments (signal-to-noise ratio, S/N),1−3 enabling the study of low concentrated systems which were not in the scope of NMR until now. NMR signal enhancement is achieved by transferring large electron polarization to surrounding nuclei via DNP. In the last decade, there has been remarkable progress in solution and solid-state DNP-NMR as well as in imaging through the application of hyperpolarization methods.4–7 As pointed out recently, this “renaissance of DNP” combined with high magnetic fields will further grow in the coming years, enabling demanding applications with better resolution.8,9 Materials and biological systems such as membrane or microcrystalline proteins,10−13 fibrils,14 ligands bound to receptors,15 polymers,16−18 surface-bound species,19−23 and other low concentrated samples were studied beneficially using DNP-NMR.13,24−26 Cryogenic temperatures are required to slow down nuclear and electron relaxation and to allow an efficient polarization transfer in solid-state DNP NMR experiments. In contrast to biological systems,15,27−29 most technically relevant materials do not show detrimental lines broadening at low temperatures.

Hence, several successful applications of solid-state DNP-NMR to materials were reported including fast and sensitive characterization of catalyst surfaces and bulk species as well as
polymers. A first example of “surface-enhanced DNP-NMR” provided by Lesage et al.,19 revealed surface bound aromatic substrates on mesoporous silicon-oxide materials at natural abundance. For this purpose, the solvent protons were first polarized by DNP, which was subsequently transferred to rare carbon nuclei. A 50-fold signal enhancement was observed. In a later work by Lelli et al., surface silicon sites were characterized using 29Si cross-polarization (CP)-MAS DNP-NMR and incipient wetness impregnation of the materials with radical-containing solvents.20 This work shows that the functional groups on catalyst surfaces can fold back rather than pointing into the core cavities. Later, Lafon et al. demonstrated the characterization of bulk silica sites buried beneath the surfaces in a similar porous material by using direct excitation 29Si DNP-NMR.21 Rossini et al. showed that the surface sites in small pores, from which the radical is excluded, can be hyperpolarized using DNP magnetization polarization transfer between the hyperpolarized nuclei on the surface and bulk regions of these nanomaterials will clarify remaining questions on their activity and function. As a result, solid-state DNP-NMR is indispensable for understanding the details of structure–function–property relationships. Previously, conventional 1H, 13C or 31P NMR studies have been performed on spherical nanoparticles and mesoporous materials, however, suffering from low sensitivity.36-50 Very recently, application of DNP enhanced MAS NMR spectroscopy on several nanoparticles has been demonstrated, showing the feasibility of the technique for studying such systems.51-53

Here we apply MAS DNP-NMR spectroscopy to investigate site-specific surface and bulk properties of pure (non-functional) as well as arginine- and lysine-functionalized silicon-oxide nanoparticles (Fig. 1).44 Precise knowledge of the surface composition and chemistry of such nanoparticles is essential for rationalizing their behavior, especially concerning their interactions with biomolecules or membranes, and so far such knowledge is still limited. The natural abundance nuclei at the molecular sites (proton, carbon, and silicon) have been characterized by DNP-NMR exploiting signal enhancements (e) of up to ~50 and ~20 for protons and silicon, respectively. The efficiency of DNP-NMR experiments depends strongly on the nanoparticle size (~10 to 100 nm), which needs to be fine-tuned for optimized DNP properties.44,54 The Q3, Q4 and Q5 types of surface silica species were characterized and surface binding of arginine and lysine were determined to be via non-covalent supramolecular interactions. These results reveal the details of the molecular structure of nanoparticles and the mechanism of polarization transfer between the hyperpolarized nuclei on the
surfaces to the bulk nuclei inside the nanoparticles. Such information is important for understanding the properties of such nanoparticles and a major aim of this article is to explore the potential of DNP-NMR to obtain additional information.

B Materials and methods

Nanoparticle synthesis

The nanoparticle synthesis is described in detail in SI 1a, ESL†. It should be noted that all nanoparticles employed were very well-defined in terms of size, with polydispersity indices well below 0.1 (see Table S1, ESL†).

Characterization of nanoparticle physical properties

The nanoparticles were characterized by means of dynamic light scattering (DLS) and transmission electron microscopy (TEM). The corresponding data are presented and explained in SI 1b and c, ESL†. It should be noted that all nanoparticles employed were very well-defined in terms of size, with polydispersity indices well below 0.1 (see Table S1, ESL†).

DNP enhanced solid-state NMR

All the DNP-NMR experiments with magic angle spinning were performed on a commercial Bruker DNP spectrometer operating at 400 MHz. A continuous wave 263 GHz microwave irradiation was generated using the Bruker gyrotron of up to several watts. A triple resonance HCN DNP probe using microwave irradiation was generated using the Bruker gyrotron.

Fig. 1. Representation of the DNP enhanced solid-state NMR spectroscopy of nanoparticles. Different nanoparticle sizes are shown with different colors and sizes of spheres. The pulse sequences used for the characterization are given below. The direct excitation and cross-polarization types of $^{13}$C/$^{29}$Si MAS NMR experiments were performed using DNP, which gives site specific information about the surface coated surfactants (arginine or lysine in the current case), and surface or bulk parts of the nanoparticle. The TEM images for the two nanoparticles with 30 and 90 nm particle sizes are shown as an example.

The DNP polarization buildup times were obtained by using 1D DNP enhanced $^{13}$C & $^{29}$Si MAS NMR spectra

The NMR spectra were recorded with $^{13}$C or $^{29}$Si CP as well as $^{29}$Si direct-excitation, by using the pulse sequences shown in Fig. 1. The spectra were recorded with and without microwave ($\mu$W) irradiation to demonstrate signal enhancement obtained by DNP (Table 1). The solid-state DNP-NMR spectra of arginine- and lysine-functionalized silica nanoparticles with a size of ~25 nm. 20 mM TOTAPOL radical was used for the DNP experiments (dissolved in 10:90% H$_2$O–D$_2$O, v:v) in samples prepared by “incipient wetness impregnation” (Fig. 2).$^{19,53}$ DNP enhancements of ~20 and ~37 were observed for silicon and proton nuclei, respectively, for the particle size of 25 nm. Such very small silica nanoparticles are interesting as they have been shown to penetrate phospholipid membranes rather easily and can be chemically modified in order to optimize such transport further and to be versatile carrier particles.$^{56}$ These values are similar to the ones reported previously.$^{19}$ However, ~30% larger enhancements were observed for direct-excitation $^{29}$Si DNP experiments recently by using a different microwave ($\mu$W) irradiation frequency, under the so-called “+” DNP condition (at $\omega_k + \omega_0$), instead of the “-” DNP condition (at $\omega_k - \omega_0$) used.$^{21}$

The power of the DNP enhancements leads to the observation of spectra with high sensitivity in a very short experiment time (~60 or ~40 seconds for the 1D CP or direct-excitation experiments, respectively, for experimental parameters see the caption of Fig. 2). This is dramatically facilitating multidimensional NMR experiments at natural abundance for site specific assignments. The spectra recorded without DNP show only very weak signals within the same experiment time. In general, three different regions in the studied nanoparticles can be distinguished: (i) the surface of the nanoparticle by applying CP $^{29}$Si NMR, (ii) the core (sub-surface) of the nanoparticle through direct-excitation $^{29}$Si NMR, and (iii) the surface bound arginine and lysine surfactant residues in the solid-state NMR experiments.

C Results and discussion

1D DNP enhanced $^{13}$C & $^{29}$Si MAS NMR spectra

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The DNP enhanced 1D $^{13}$C CP and $^{29}$Si direct-excitation/CP NMR spectra of arginine- (A) and lysine- (B) functionalized silicon-oxide nanoparticles with ~25 nm particle size. The spectra recorded with and without DNP are shown in black and grey colors, respectively, at 8 (for $^{29}$Si) or 12 (for $^{13}$C) kHz MAS and ~100 K. The $^{13}$C CP, $^{29}$Si direct-excitation and $^{29}$Si CP NMR spectra were recorded with 1024, 32 and 4 scans, respectively, similarly for the $^{13}$C and $^{29}$Si CP NMR experiments. Relaxation delays of 2 and 10 seconds were used for the CP and direct-excitation type experiments, respectively. Contact times of 0.4 and 3 ms for the $^{13}$C and $^{29}$Si CP NMR experiments. The carbon signals from residual TEOS are indicated by asterisks.

Fig. 2

The $^{29}$Si CP spectra show different Q-type ($Q_n = \text{Si} (\text{OSi})_n(\text{OH})_{4-n}$) silicon sites for both coated and pure silica materials, which are observed at $\delta_{^{29}\text{Si}} = \sim 110$, $\delta_{^{29}\text{Si}} = \sim 100$, and $\delta_{^{29}\text{Si}} = \sim 90$ ppm (see SI 2, ESI!) for 1D $^{13}$C and $^{29}$Si spectra of non-functional silicon-oxide nanoparticles. The amount of different silicon sites are determined by integrating the specific resonances in spectra recorded via CP (represented below in terms of ratios, %, in each individual spectrum. The differences in those individual ratios between different samples are compared); which are 10/8 ($Q_2$), 57/71 ($Q_3$), and 33/21 ($Q_4$) % for the 25/100 nm nanoparticles coated with arginine, respectively (the ratios and some of the fits are in SI 3, ESI, as well as for other particles). A remarkable observation is that no $Q_2$ type silicon site was detected in conventional $^{29}$Si MAS NMR experiments for similar nanoparticles studied previously, but only the $Q_3$ and $Q_4$ sites were identified. The $Q_2$ type silicon sites are determined to be ~10% in CP based spectra (Fig. 2), which clearly proves the power of the DNP-NMR technique in the determination of chemically different sites on surfaces at low concentrations and natural abundance. No T-type silicon sites are observed (T$_n$ = Si R(OX)$_3$...n(OSi)$_m$ surface silicon sites with a chemically bound functional group; R and X are organic functional sites) using $^{29}$Si CP DNP-NMR experiments (Fig. 2A and B), indicating that the arginine/lysine binds to the nanoparticle surface with supramolecular interactions such as hydrogen bonding or electrostatic forces, but not via chemical bonds as observed for other mesoporous silica materials.

An interesting behavior appears when the CP (indirect DNP) and direct-excitation (direct DNP) $^{29}$Si spectra are compared (Fig. 2). All three silicon sites ($Q_2$–$Q_4$), are polarized via CP ($e_{Q_2} = \sim 35/3$, $e_{Q_3} = \sim 39/4$, and $e_{Q_4} = \sim 35/5$ for the 25/100 nm nanoparticles, respectively). Direct-excitation, however, results in spectra with dominating $Q_4$ type of silicon signals with high enhancement. Whereas, the $Q_2$ and $Q_3$ signals are present in the direct excitation $^{29}$Si spectra with less intensity due to a less efficient polarization with much lower enhancements compared to $Q_4$ enhancements (Table 1). This difference between the direct-excitation and CP $^{29}$Si DNP experiments has been reported previously for a mesoporous silica material, which shows that the surface silicon sites ($Q_2$–$Q_3$) are best polarized by CP, whereas, the sub-surface sites ($Q_4$) are polarized by direct-excitation, which is also the case for the studied nanoparticles. As a result, it can be concluded that there is a preferential DNP polarization phenomenon in both silica based mesoporous and spherical materials, which leads to different DNP enhancements for different $^{29}$Si sites, and as a result, different signal intensities in the spectra. This observation is difficult to rationalize since the $Q_2$ and $Q_3$ sites are very close to the surface or they are on the surface, and as a result they are closer to the radicals compared to $Q_4$. However, because of a possible larger amount of $Q_4$ nuclei present in the polarizable region of the nanoparticles, the signals appear much more intense than the signals of other sites in the $^{29}$Si MAS NMR spectrum. The localization of the radical on the nanoparticle surface can also lead to the bleaching of silicon signals close to the radical, which might result in the observed less intense $Q_2$ and $Q_3$ signals. The $^{29}$Si MAS NMR spectra recorded without $\mu$W irradiation are similar in signal intensity, indicating the absence of any significant bleaching effect.

Determination of the absolute DNP enhancements were done on one of the nanoparticles, to estimate the real signal enhancement by DNP NMR at ~100 K with radicals compared...
to a sample without radicals and measured at ambient temperature. For this purpose, 11 nm arginine surface functional nanoparticles were used. The comparison of the μW-on $^{29}$Si CPMAS and MAS spectra recorded at 100 K to the μW-off spectra recorded at 100 K resulted in DNP enhancements of 43 ($^1$H DNP) and 14 ($^{29}$Si DNP) for this specific nanoparticle preparation. And finally, for the determination of absolute DNP enhancements, we compared the μW-on $^{29}$Si CPMAS and MAS spectra recorded at 100 K to the spectra recorded at 280 K but without radicals. The results show that the DNP enhancements are 18 ($^1$H DNP) and 26 ($^{29}$Si DNP), which shows that the absolute $^1$H DNP enhancement is nearly half of the direct-comparison (μW on/off) value, whereas, the $^{29}$Si DNP enhancement is nearly two times larger indicating the favored shorter relaxation times due to the presence of radicals (SI 4, ESI†). Similar comparisons were done on other silica materials as well.$^{52,57}$

The effect of nanoparticle size & composition of Si-sites

After determination of the enhancement of each type of signal, we investigated the nanoparticle-size-dependent changes in the surface/sub-surface silicon hyperpolarization ratios by comparing the signals observed from the DNP enhanced $^{29}$Si CP spectra particles with different sizes applying 5 and 20 mM radicals (SI 3, ESI†). The integral of the whole spectrum is normalized to 100%, to be able to compare the signal intensity changes among different spectra. In the 25 nm particle coated with arginine, the signals of Q$_2$, Q$_3$, and Q$_4$ silicon sites occur with 10, 57 and 33%, respectively, as determined using integration of the CP spectra obtained from the sample containing 20 mM radicals.

The same sample prepared without any radicals, however, shows slightly different silicon ratios of 8, 67, and 25% for the signals of same sites. The Q$_3$ signals become more abundant and the Q$_4$ signals become less abundant for the samples prepared without radicals, which indicates that the DNP measurements favor the detection of Q$_2$ signals. For the 90 nm nanoparticles containing 20 mM radicals, the Q$_4$ sites (42%) are slightly more abundant compared to Q$_2$ (4%) and Q$_3$ sites (54%). Interestingly, 90 nm particles containing 5 mM radicals show an increased amount of Q$_2$ signal (12%), and less intense Q$_3$ and Q$_4$ signals (50 and 38%, respectively), which indicates that a high radical content for the large particles can cause removal of the DNP-NMR signals from the Q$_2$ and Q$_3$ surface species.

The direct excitation $^{29}$Si measurements on the 90 nm particles with 20 mM shows significantly different ratios for different silicon signals compared to the CP spectra. The Q$_4$ signals are present in a much larger ratio (~83%). This is also due to the observation of an additional broader Q$_4$ type signal (Q$_{4\text{m}}$) by direct excitation (see Fig. 2). However, the Q$_2$ and Q$_3$ signals are much less intense (~1 and 16%, respectively) compared to the CP experiments. The experiments using the sample containing 5 mM radicals show that the Q$_3$ signal is observed with ~1 to 2% and the Q$_1$ signal is increased to ~26%, whereas, the Q$_4$ signal decreases to ~73%. These results once again indicate that the Q$_3$ sites are buried beneath the surface and their signals are less bleached out in the presence of a large radical content, whereas, the signals of Q$_3$ and Q$_4$ sites are more sensitive to the radical concentration since they appear either on the surface or close to it and are in closer contact with the radicals. The use of longer relaxation delays for the $^{29}$Si direct-excitation experiments, such as 50 or 1000 seconds instead of 10, results in an increase of the narrow Q$_4$ component, whereas, the broad Q$_2$ signal appears to be reduced (SI 5, ESI†), as a result of different relaxation behavior of these two components. We did not observe a relaxation-delay dependent enhancement change.

The $^{13}$C CP spectra recorded for the arginine and lysine functionalized nanoparticles are shown in Fig. 2 (the same spectrum for the pure sample without surface coating is shown in SI 2, ESI†). With the DNP enhanced spectra (shown in black in Fig. 2), very strong signals are observed, which are absent in the NMR spectra recorded without μW (shown in grey in Fig. 2). The estimated DNP enhancement is ~40 to 50 for these experiments, without which the NMR signal observation is practically not possible. Signals from the surface-bound arginine and lysine species are clearly seen in the spectra. Moreover, two additional carbon signals were observed, similar to previous studies on mesoporous materials,$^{19}$ which originate from incomplete condensation of the starting material, tetraethylorthosilicate (TEOS), due to fast nucleation of the silica primary particles.$^{18}$ The carbonyl and C$_5$ signals appear weaker due to the short CP contact time which was optimized for the detection of aliphatic signals.

The effect of the nanoparticle size and DNP efficiency

The nanoparticle size has a remarkable effect on the observed DNP enhancements (Fig. 3 and 4 and Table 1). The enhancements decrease by an order of magnitude upon increasing the

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**Fig. 3** The effect of the silicon-oxide nanoparticle size on the DNP enhancements $\epsilon_{\text{DNP}}$ for the particles with diameters of (A) 100, (B) 90, (C) 50, (D) 25 nm. 10 $^{25}$Si CPMAS spectra of arginine functionalized silicon-oxide nanoparticles are recorded with (black, 32 scans) and without (grey, 128 scans) μW irradiation to quantify the proton-DNP enhancements. The spectra were recorded at ~100 K, 9.4 T, 8 kHz MAS, and by using samples with a 20 mM TOTAPOL concentration in zirconium oxide rotors.
The data shown are for the arginine coated for 10–90 nm particles, whereas for a particle size from ~10 nm (e.g., $e^\text{CP}_\text{Si} = 43$ and $e^\text{MAS}_\text{Si-Q}_4 = 21$) up to ~100 nm (e.g., $e^\text{CP}_\text{Si} = 4$ and $e^\text{MAS}_\text{Si-Q}_4 = 3$) in direct-excitation and CP based silicon spectra, respectively. A larger particle size results in a decreased DNP efficiency and a lower enhancement. This phenomenon can be described as follows: All silicon sites can be excited by direct-excitation (core and the surface sites) NMR; however, only a fraction of the atoms can be polarized in DNP-NMR experiments due to limited DNP penetration depth, which in our case can be estimated to be ~4.2 or ~5.7 nm (see text below). Consequently, larger than a certain nanoparticle size, there can be a region in the core of the nanoparticle which cannot be polarized but which can be still detected. This decreases the $\text{Si}_{\text{polarized}}/\text{Si}_{\text{non-polarized}}$ ratio, and as a result decreases the apparent DNP enhancements. The direct-excitation silicon experiments show a maximum of DNP enhancements at particle sizes between ~13 and 25 nm (Fig. 4). Our data show that, for the $^{29}\text{Si}$ direct-excitation experiments, the penetration depth is not sufficient to polarize all the nuclei above a particle size of ~25 nm. The observed limited DNP penetration depth is also due to a relatively less efficient silicon spin diffusion attributed to the presence of an extended Si–O network composed of unlabelled silicon nuclei which also lacks protons, compared to a fully protonated and uniformly carbon labeled protein which has faster spin diffusion. As control experiments, the samples were prepared by using appropriate amounts of radical containing solutions to be able to be sure about the radical concentration on the surfaces of nanoparticles with different surface areas. The results show a similar size-enhancement profile as shown in Fig. 4 (data not shown).

**Estimation of DNP penetration depth**

Similar to our “DNP penetration depth” argument, a “spin diffusion barrier radius” was determined in a recent work by Lafon et al., which describes the extent to which the sub-surface nuclei can be polarized by the radical at the surface of the particle. A “spin diffusion barrier radius” was estimated to be ~35 Å for a mesoporous silica material with an average pore wall thickness of 15 Å. Under these conditions all signals were determined to originate from core nuclei and DNP polarization transfer was proposed to be a direct way. However, due to the larger size of our particles the effect of spin diffusion is sizeable for both silicon and proton nuclei, and there should be a separation of core/bulk nuclei.

In a study performed on 100–200 nm thick GNNQNY peptide crystals by van der Wel et al., only a slight decrease (~25%) in the proton DNP enhancement was observed when the enhancement of the radical containing solvent and the isolated protein enhancements were compared. The solvent signals show the maximum proton enhancement since the radical polarizes this part first. On the other hand, the protein signals represent an average enhancement since the hyperpolarization diffuses through the crystal, and as a result a polarization gradient occurs with simultaneous detection of highly polarized nuclei which are close to the solvent and less polarized nuclei which are in the core of crystal. Moreover, the effect of spin diffusion is explicitly represented in this work, and it was shown that larger spin diffusion constant leads to a less dramatic DNP enhancement deterioration in a thick sample. We observed a much larger effect of the particle size on the DNP enhancement, which could be a result of much faster spin diffusion in a uniformly labeled and protonated protein crystal, in contrast to our less-proton containing and natural abundance silica nanoparticles. This observation emphasizes the need for a careful optimization of the particle size for obtaining maximum sensitivity.

Moreover, a recent hyperpolarization study was performed on polycrystalline diamond samples by utilizing natural defects as an electron source, as well as hyperpolarized silicon nano/micro particles for various purposes including MRI. In the former study by Casabianca et al., polycrystalline diamond samples containing naturally occurring nitrogen-defects as electron sources were studied for different particle sizes and the resulting DNP properties were determined. It has been found that hyperpolarization was only successful for the diamond samples with larger particle sizes, >100 to 300 nm up to microns. Enhancements of up to 15–20 were observed at ~2 K. Our results indicate an opposite trend, most probably due to our approach involving exogenous radicals in comparison to their approach utilizing intrinsic defects as electron sources.
In the latter study by Aptekar et al., large silicon nano-micro particles (from 100 nm up to several microns) were similarly hyperpolarized at ~4 K leading to sensitive MRI applications. Compared to these studies, our work is unique in the sense of achieving hyperpolarization for much smaller nanoparticles of 10–100 nm, which can lead to more efficient \textit{in vivo} imaging or molecular targeting applications.

In order to obtain further information on the effect of nanoparticle size to the solvent surrounding the nanoparticles, direct excitation $^1$H MAS DNP-NMR spectra were recorded. By monitoring the changes in the observed DNP enhancements from the $^1$H direct excitation and $^{29}$Si CP NMR experiments, the polarization behavior of the solvent ($\text{H}_2\text{O}–\text{D}_2\text{O}, 10:90\%$ v:v) and the nanoparticle are determined separately (Fig. 4B).

A decrease in proton DNP enhancements is observed with increasing nanoparticle size in the $^{29}$Si CPMAS experiment, which is explained above by a DNP penetration depth effect. Similarly, a decrease in the proton DNP enhancement of the solvent is then observed with increasing particle size, which cannot be explained with the DNP penetration depth effect due to the readily available radicals in the solvent. In this case, the unfavorably increased radical content in the solvent part of the system might have caused the observation of reduced enhancements for larger particles. We have similarly shown that the $^1$H DNP enhancement of a uniformly labeled proline sample dissolved in a glycerol–water solvent decreases when the radical concentration is unfavorably increased (above ~25 mM TOTAPOL).

For a control experiment, we performed DNP-NMR experiments on a 90 nm coated particle by using a four-fold lower radical concentration of 5 mM (Table 1). As expected, reducing radical concentration results in slightly lower $^{13}$C MAS enhancements for the silicon sites compared to the 20 mM sample. The TEO$_6$$^{13}$C$_2$ signal, which is inside the nanoparticle, is decreased from 9 to 6–7. Interestingly, lowering radical concentration increases the DNP enhancements detected directly by proton direct-excitation NMR and indirectly by silicon CP NMR (from 24 to 32 for the direct and 6 to 8 for the indirect NMR, respectively). This suggests that, a radical concentration of 20 mM is unfavorably high for the surface species as well as for the solvent for the 90 nm particle size.

\textbf{A model calculation for determining DNP penetration depth}

A model calculation was performed to analyze the effect of nanoparticle size on the DNP enhancement and to estimate the extent of polarization transfer from the radical into the core regions of the particles (the polarization penetration depth). The total surface area (Fig. 5, black solid line) of the nanoparticles per rotor volume decreases by using larger nanoparticles which, as a result, changes the number of TOTAPOL molecules per unit surface area. In the simplified model, an ~7- to 8-fold increase in the apparent radical concentration per unit surface area was observed by increasing the nanoparticle size from ~13 to ~100 nm (see SI 7, ESI$^\dagger$ for the detailed description of the calculation). The DNP enhancements are obtained from the CP (red solid line, via $Q_3$ signal) or direct-excitation (blue solid line, via $Q_4$ signal) based $^{29}$Si DNP-NMR spectra, (Fig. 5). The dotted lines represent the calculated DNP enhancements for each nanoparticle size which takes the ratio of the polarizable and non-polarizable parts of the nanoparticles into account by using the formula shown in SI 7, ESI$^\dagger$ (represented in Fig. 5D). In the ideal case of a sharp border between the polarized and non-polarized regions, the DNP polarization penetration depth is estimated to be ~4.2 nm when using the enhancement values from CP-based $^{29}$Si experiments and ~5.7 nm when using enhancements from direct-excitation-based $^{29}$Si experiments. These values are similar to the ones reported for the mesoporous silica particles. However they are much smaller than the values obtained from proton-mediated transfer on a fully labeled and protonated protein. One important point to note here is the presence of residual TEO$_6$ species in the nanoparticle, whose carbon signals are observed in $^{13}$C CPMAS spectra shown in Fig. 2. The exact position of these moieties is unknown, but a random distribution in the particle core and close to surface is possible. The presence of an additional residual proton bath could have facilitated the polarization transfer.
from the electron to the nanoparticle core throughout the penetration depth of ~4.2 nm distance (via spin-diffusion), which exceeds the transfer range of a $^1$H–$^{29}$Si CP.

**DNP polarization build-up time**

DNP polarization transfer from the electrons of the radical to the surrounding nuclei has a characteristic polarization build-up behavior with a time constant $\tau_B$. The magnitude of this time constant can tell about the nature of this process. We determined the polarization build-up times for proton and silicon nuclei by using DNP enhanced $^{29}$Si cross-polarization and direct-excitation NMR experiments, respectively (the results are given in Table 1 and shown in Fig. 6). First, we observed two different components in the buildup curves of the Q$_4$ type silicon signals (30% and 70% population, respectively), which were determined by using saturation-recovery type 1D $^{29}$Si MAS DNP-NMR spectra with $\mu$W irradiation ($^{29}$Si-Q$_4^B$ and $^{29}$Si-Q$_4^B$). In a recent work by Lafon et al. a similar situation is observed, in which the presence of two different components was explained by the presence of two distinct Q$_4$ silicon sites which are either close to or remote from TOTAPOL. In addition to this, we observed a strong dependence of $\tau_B$ on the nanoparticle size. With increasing particle size, the polarization builds up much faster and as a result $\tau_B$ decreases rapidly ($\tau_{B-Fast} = 66/11$ s and $\tau_{B-Slow} = 981/613$ s for the 13/100 nm nanoparticles, respectively). As we discussed before, an increase in the particle size results in an increase in the TOTAPOL per-unit-surface-area, and this leads to a faster build-up of the silicon magnetization. The very long build-up times are a result of long longitudinal relaxation times; however, the effect of slow spin diffusion can also contribute to this effect for the very large spherical particles studied which hinders the incorporation of TOTAPOL beneath the surface. On the other hand, a reduced radical concentration slows down the polarization build-up of silicon signals originating from sub-surface regions, and the $^{29}$Si-Q$_4^B$ for the slow component changes from 626 (for 20 mM) to 1036 seconds (for 5 mM).

Narrower silicon lines are observed in 1D $^{29}$Si MAS DNP-NMR spectra when longer DNP polarization times are used, similar to the situation observed for the mesoporous materials (see SI 3, ESI$^\dagger$ for the ratios of different silicon sites and SI 5, ESI$^\dagger$ for the linewidth changes for Q$_4$ site by increased polarization time). This can be explained by an increase of slow building-up (narrow) of Q$_4$ signals at longer polarization times compared to the fast building-up (broad) one. To quantify this effect, the ratios of the broad and narrow components of the Q$_4$ sites were determined with the $^{29}$Si MAS experiments recorded with 10 and 1000 seconds of polarization times for the 90 nm particle (SI 3, ESI$^\dagger$). The broad component decreases from 46 to 27% with increasing polarization time, whereas the narrower component increases from 37 up to 63%.

**D Conclusions**

In conclusion, we present an application of dynamic nuclear polarization-enhanced solid-state NMR to pure and amino acid functionalized silica nanoparticles of different sizes and low polydispersity. This method allows the ultra-fast and ultra-sensitive characterization of pure and arginine/lysine modified silica particles, where such functionalization is important for potential further modification of such nanoparticles and in general for their applications. The surface, core (sub-surface) and the surface-attached species could be probed separately and unique structural information regarding the structure as a function of the depth from the surface could be obtained. Three different Q-type silicon sites (Q$_2$, Q$_3$, Q$_4$) are determined in the spectra of silica particles. No T-type sites were observed, which indicates an arginine/lysine binding to occur via either hydrogen-bonding or electrostatic forces. The DNP properties depend in a remarkable fashion on the particle radius. An enhancement increase of nearly an order of magnitude was observed by decreasing the particle size from 100 nm down to 13 nm. Surprisingly, the increased particle size resulted in a faster $^{29}$Si DNP polarization buildup time, due to increased surface radical concentration compared to smaller nanoparticles. Different silicon sites were polarized to different extents by performing CP or direct-excitation based experiments. A DNP penetration depth (or in other words...
spin diffusion barrier) of ~4.2/5.7 nm (via $^{29}$Si CP/direct-excitation experiments) was estimated by using the particle size dependent DNP enhancements.

The DNP-NMR studies on silica nanoparticles reveal details about polarization efficiency and transfer depending on the particle size. We believe that this knowledge will improve our structural understanding of nanoparticles and DNP processes substantially, and will open up new research directions. In particular, unique information regarding the surface composition and chemistry of nanoparticles becomes accessible using this method, which is essential for the understanding of the structure–function relationship. Only based on such detailed surface structural information rational further developments of nanoparticle functionalization is possible and the DNP-NMR method offers here unique additional insights. Based on such information more efficient delivery systems or catalytically active nanoparticles may be constructed. Furthermore, the efficient hyperpolarization of small silica nanoparticles with presented efficiency can be utilized in magnetic resonance imaging applications.

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Notes and references


