



3-Iodothyronamine Activates a Set of Membrane Proteins in Murine Hypothalamic Cell Lines

Julia Bräunig^{1,2}, Stefan Mergler³, Sabine Jyrch^{1,2}, Carolin S. Hoefig^{4,5}, Mark Rosowski⁶, Jens Mittag^{5,7}, Heike Biebermann^{1,2} and Noushafarin Khajavi^{1,2*}

¹ Charité – Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany, ² Institute of Experimental Pediatric Endocrinology, Berlin, Germany, ³ Klinik für Augenheilkunde, Charité – Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany, ⁴ Institute of Experimental Endocrinology, Charité – Universitätsmedizin Berlin, Corporate Member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Berlin, Germany, ⁵ Department of Cell & Molecular Biology, Karolinska Institutet, Stockholm, Sweden, ⁶ Department Medical Biotechnology, Institute of Biotechnology, Technical University of Berlin, Berlin, Germany, ⁷ University of Lübeck – Center of Brain Behavior and Metabolism, Lübeck, Germany

OPEN ACCESS

Edited by:

Ichiro Maruyama,
Okina Institute of Science and
Technology, Japan

Reviewed by:

Rocco Bruno,
Independent researcher, Matera, Italy
Marco António Campinho,
Centro de Ciências do Mar (CCMAR),
Portugal

Tania M. Ortiga-Carvalho,
Universidade Federal do Rio de
Janeiro, Brazil

*Correspondence:

Noushafarin Khajavi
noushafarin.khajavi@charite.de

Specialty section:

This article was submitted to
Molecular and Structural
Endocrinology,
a section of the journal
Frontiers in Endocrinology

Received: 18 June 2018

Accepted: 21 August 2018

Published: 11 September 2018

Citation:

Bräunig J, Mergler S, Jyrch S,
Hoefig CS, Rosowski M, Mittag J,
Biebermann H and Khajavi N (2018)
3-Iodothyronamine Activates a Set of
Membrane Proteins in Murine
Hypothalamic Cell Lines.
Front. Endocrinol. 9:523.
doi: 10.3389/fendo.2018.00523

3-Iodothyronamine (3-T₁AM) is an endogenous thyroid hormone metabolite. The profound pharmacological effects of 3-T₁AM on energy metabolism and thermal homeostasis have raised interest to elucidate its signaling properties in tissues that pertain to metabolic regulation and thermogenesis. Previous studies identified G protein-coupled receptors (GPCRs) and transient receptor potential channels (TRPs) as targets of 3-T₁AM in different cell types. These two superfamilies of membrane proteins are largely expressed in tissue which influences energy balance and metabolism. As the first indication that 3-T₁AM virtually modulates the function of the neurons in hypothalamus, we observed that intraperitoneal administration of 50 mg/kg bodyweight of 3-T₁AM significantly increased the c-FOS activation in the paraventricular nucleus (PVN) of C57BL/6 mice. To elucidate the underlying mechanism behind this 3-T₁AM-induced signalosome, we used three different murine hypothalamic cell lines, which are all known to express PVN markers, GT1-7, mHypoE-N39 (N39) and mHypoE-N41 (N41). Various aminergic GPCRs, which are the known targets of 3-T₁AM, as well as numerous members of TRP channel superfamily, are expressed in these cell lines. Effects of 3-T₁AM on activation of GPCRs were tested for the two major signaling pathways, the action of G_{α_s}/adenylyl cyclase and G_{i/o}. Here, we demonstrated that this thyroid hormone metabolite has no significant effect on G_{i/o} signaling and only a minor effect on the G_{α_s}/adenylyl cyclase pathway, despite the expression of known GPCR targets of 3-T₁AM. Next, to test for other potential mechanisms involved in 3-T₁AM-induced c-FOS activation in PVN, we evaluated the effect of 3-T₁AM on the intracellular Ca²⁺ concentration and whole-cell currents. The fluorescence-optic measurements showed a significant increase of intracellular Ca²⁺ concentration in the three cell lines in the presence of 10 μM 3-T₁AM. Furthermore, this thyroid hormone metabolite led to an increase of whole-cell currents in N41 cells. Interestingly, the TRPM8 selective inhibitor (10 μM AMTB) reduced the 3-T₁AM stimulatory effects on cytosolic Ca²⁺ and whole-cell

currents. Our results suggest that the profound pharmacological effects of 3-T₁AM on selected brain nuclei of murine hypothalamus, which are known to be involved in energy metabolism and thermoregulation, might be partially attributable to TRP channel activation in hypothalamic cells.

Keywords: 3-T₁AM, signaling, hypothalamus, GPCR, TRP Channel

INTRODUCTION

3-iodothyronamine (3-T₁AM) is a decarboxylated and deiodinated derivative of thyroid hormones (1, 2). Although several studies detected 3-T₁AM in human blood (3, 4), the mechanism of physiological action of this compound in the human body remains undefined. Administration of 3-T₁AM in rodents blocks the hypothalamic–pituitary–thyroid axis and results in concentration-dependent reversible effects on body temperature, energy metabolism, cardiac and neurological functions (1, 5). Previous observations in rodents demonstrated the accumulation of 3-T₁AM in different tissues such as kidney, liver, muscles, and brain (6). In mice, after administration of a radioisotope labeled [¹²⁵I]-3-T₁AM, ¹²⁵I was detected in the brain (6). In rats, site-directed injections of 3-T₁AM into the locus coeruleus elicits significant neuronal firing rate changes in the selected brain nuclei such as the paraventricular nucleus (PVN) of the hypothalamus (7). Interestingly, the target areas of 3-T₁AM in the brain nuclei are mostly involved in energy homeostasis, arousal, and memory retrieval (8–11). Presumably, different effects of 3-T₁AM, such as anaptyrexia and food consumption might be centrally mediated via the hypothalamus (12–14). Due to the profound pharmacological effects of 3-T₁AM and its accumulation in the selected tissue, numerous studies over the last years have been devoted to investigating the signaling property of this thyroid hormone metabolite.

The first target of 3-T₁AM is the trace amine associated receptor 1 (TAAR1), a trace amine-activated G protein-coupled receptor (GPCR) (1). 3-T₁AM induces G_α_s/adenylyl cyclase signaling in rodent TAAR1 and human TAAR1-transfected cells (1). Additionally, different studies described several other GPCRs as 3-T₁AM targets, mainly *in vitro* and in overexpressing systems. These GPCRs belong to the group of aminergic GPCRs (15) such as the α-2A-adrenergic receptor (ADRA2A (16)), the β₂-adrenergic receptor (ADRB2) (17), the muscarinic receptor 3 (CHRM3) (18), or the serotonin receptor 1b (5-HT1b) (19). Moreover, 3-T₁AM modulates calcium and potassium homeostasis through an intracellular calcium channel, known as “ryanodine receptor” in adult rat cardiomyocytes (20).

Recent studies identified non-selective cation channels such as the transient receptor potential channel melastatin 8 (TRPM8) and the transient receptor potential vanilloid 1 (TRPV1) as novel targets of 3-T₁AM (21–23). Classically, TRPM8 is known as a cold and menthol receptor and is a temperature-sensitive receptor in excitable cells (24). Its activation induces a depolarization of the cell membrane leading to action potentials. The same function principle applies to TRPV1, which is known as a heat- and capsaicin receptor (25). Together, these properties implicate these TRPs as possible transducers of cold or warm

stimuli not only within the hypothalamus (26), but also in keratinocytes of human skin (27) and neurons on human corneal nerve fibers (28, 29).

Different studies demonstrated that TRPs are the major downstream effectors of GPCRs and the signaling cascades that emanate from the activation of GPCR evoke TRP channel activity (30, 31). There is a wide distribution of TRPs in tissues that influence energy homeostasis and thermoregulation. Expression of TRPs in various tissues such as hypothalamus, peripheral sensory neurons, gastrointestinal tract, liver, adipocytes, and ocular tissues strongly suggest the possible role these ion channels play in energy balance and metabolism as well as thermoregulation (32–37). Modulation of TRPs via 3-T₁AM raises the question of what could be the 3-T₁AM-induced signalosome and whether there is a link between stimulatory effects of 3-T₁AM in tissues that pertain to metabolic- and/or thermo-regulation and TRPs.

Here, we identified the stimulatory effect of 3-T₁AM in murine hypothalamic nuclei and explored the underlying mechanism behind this effect in murine hypothalamic cell lines. The results of this study show a stimulatory effect of 3-T₁AM on Ca²⁺ mobilization and whole-cell currents in murine hypothalamic cells and that this effect is associated with TRPM8 activation.

METHODS

Mice Experiments Immunohistochemistry

In collaboration with the Karolinska Institute, Sweden, C57BL/6J mice (4 in each group) were i.p. injected with 50 mg/kg body weight 3-T₁AM solved in 60% DMSO and 40% PBS, control mice with 60% DMSO and 40% PBS (volume of injection was 5 μl/g body weight). After 60 min, animals were transcardially perfused with PBS and 10% formalin (European Community Council Directives (86/609/EEC) and approved by Stockholm's Norra Djurförsöksetiska Nämnd). Fixated murine brains were successively incubated in 10, 20, and 30% sucrose solution over several days. Brains were cut at a cryotom into 30 μm slices and placed in a 48 well plate filled with PBS. Slices were blocked with a blocking buffer (TBS, 0.25% gelatin from porcine skin and 0.5% triton X100) for 2 h, subsequently incubated with a c-FOS antibody, rabbit anti mouse (1:200; Santa Cruz Biotechnology, Santa Cruz, CA, USA) over night at 4°C and finally with an Alexa Fluor 549 antibody, goat anti rabbit (1:200; Jackson ImmunoResearch) for 2 h at room temperature. Between each step, the slides were washed 3 × 1 min with TBS (tris-buffered saline). The brain slices were placed on

glass slides and mounted with VectaShield containing DAPI (Vector Laboratories, Burlingame, CA, USA). Pictures were taken with a Keyence BZ-9000 microscope (20× magnification) and optimized with the ImageJ software. C-FOS positive cells were counted with the ImageJ software using two brain slides of each animal for each brain loci.

Cell Culture and mRNA Isolation

GT1-7 (mouse hypothalamic gonadotropin-releasing-hormone neuronal cell line) were purchased from MERK, (38). mHypoE-N39 (N39) and mHypoE-N41 (N41), both embryonic mouse hypothalamic cell lines, were acquired from Cedarlane, established by Belsham et al. (39). A screening profile of neuronal markers of all three cell lines can be viewed here (<https://www.cedarlanelabs.com/Products/Listing/Hypothalamic>). The cells were cultured in Dulbecco's modified Eagle's medium (DMEM, Biochrom GmbH, Berlin, Germany), supplemented with 10% fetal calf serum (FCS) and 1% penicillium/streptavidin at 37°C in humidified air containing 5% CO₂.

Cells were seeded in T75 flask and grown to 80% confluence. Cells were harvested in three different passages, spanning from passage three to passage six. Total mRNA was isolated by chloroform/phenol extraction, a DNase digestion was performed and samples were stored at -80°C.

Quantitative PCR to Determine GPCR and TRP Channel Expression Profiles

A quantitative PCR (qPCR) was performed to determine the expression level of several GPCRs and TRPs. QPCR primers including their efficiency are listed in **Supplemental Table 1**. *Pgk1* was chosen as the reference gene, as previously recommended (40). First, mRNA was transcribed into cDNA by the Omiscript RT kit (Qiagen) using random hexamers (Applied Biosystems) and Oligo dTs (Promega, Madison, USA). Absolute QPCR Mix, SYBR Green, no Rox (Thermo Scientific, Germany) was used for qPCR on a Stratagene Mx3000P System using 100 nM per primer. PCR reaction underwent an initial cycle at 95°C for 15 min followed by 42 cycles at 95°C for 15s, primer specific annealing step 60°C for 30s and an elongation step 72°C for 45s, and elongation at 72°C for 7 min and finally temperature holding at 4°C. Melting curve analysis was performed to confirm the specificity of the PCR reaction. Data was processed using the ΔCt method. We used the slope of a standard curve to determine the amplification efficiency for each primer pair (efficiency = $10^{-1/\text{slope}}$). Each of the three passages of every cell line was measured in duplicates together with a sample without reverse transcription to exclude genomic DNA contamination.

G α_s and G i/o Signaling of Endogenous Expressed GPCRs in Hypothalamic Cell Lines

G α_s and G i/o signaling were determined by measuring cAMP accumulation using the AlphaScreen technology (PerkinElmer Life Science, Boston, MA, USA) as previously described (15). Cells were cultured in poly-L-lysine (Biochrom GmbH,

Berlin, Germany) coated 96-well plates (1×10^{-4} cells / well). Seventy-two hours after seeding, stimulation was performed by means of using a stimulation buffer (138 mM NaCl, 6 mM KCl, 1 mM [MgCl₂*6H₂O], 5.5 mM glucose, 20 mM HEPES, 1 mM [CaCl₂*2H₂O], 1 mM IBMX). For G α_s signaling, cells were incubated for 45 min with either 3-T₁AM (Santa Cruz Biotechnology, Dallas, TX, USA), serotonin (5-HT, Sigma-Aldrich, St. Louis, MO, USA), norepinephrine (NorEpi, Sigma-Aldrich, St. Louis, MO, USA), isoproterenol (ISOP, Sigma-Aldrich, St. Louis, MO, USA), or phenethylamine (PEA, Sigma-Aldrich, St. Louis, MO, USA) in a concentration of 10 μ M or only stimulation buffer to monitor the basal cAMP content. 3-T₁AM was diluted from a 10 mM stock solution using DMSO as solvent. H₂O was used as solvent for serotonin and norepinephrine and PBS with 0.1% BSA was used as the solvent for isoproterenol and phenethylamine. For G i/o pathway examination, cells were additionally stimulated with 50 μ M forskolin (FSK, AppliChem GmbH, Darmstadt, Germany) to activate the adenylyl cyclase for a total of 45 min. Afterwards, cells were lysed at 4°C on a shaking platform. Intracellular cAMP accumulation was determined by a competitive immunoassay based on the AlphaScreen assay kit according to the manufacturer's instructions and measured using a Berthold Microplate Reader (Berthold Technologies GmbH & Co. KG, Bad Wildbad, Germany). Cyclic AMP concentrations were normalized to protein contents, which was measured with the Pierce BCA Protein Assay Kit (Thermo Scientific, Germany).

Determination of Intracellular Ca²⁺ Concentration

To monitor time-dependent changes in intracellular free Ca²⁺ levels ([Ca²⁺]_i) in single-cells, cells were pre-incubated with culture medium containing fura-2/AM (2 μ M) for ~30 min at 37°C. Loading was stopped with a Ringer-like (control) solution containing: 150 mM NaCl, 6 mM CsCl, 1 mM MgCl₂, 10 mM glucose, 10 mM HEPES, and 1.5 mM CaCl₂ at pH 7.4. Where a blocker was used, pre-incubation was performed 30 min before the measurement. Fluorescence measurements were performed on the stage of an invert microscope (Olympus BW50WI) and a camera (Olympus XM-10) in connection with a LED-Hub (Omikron, Rodgau-Dudenhoven, Germany). Fura-2/AM fluorescence was excited at 340 and 380 nm alternatingly and emission was detected from small cell clusters every 4 s at 510 nm. Results are shown as mean traces of $f_{340\text{nm}}/f_{380\text{nm}} \pm \text{SEM}$. Drugs were dissolved in dimethyl sulfoxide (DMSO) to obtain a stock solution and diluted in Ringer-like solution to obtain a working concentration which did not exceed 0.1%. For image acquisition and data evaluation, the life science imaging software cellSens was used (Olympus, Hamburg, Germany).

Planar Patch-Clamp Recordings

For electrophysiological recordings, the semi-automated planar patch-clamp technique was used as previously described (41). Whole-cell currents were evaluated in conjunction with an EPC10 amplifier and PatchMaster acquisition software (HEKA, Lambrecht, Germany) as well as PatchControl software (Nanon,

Munich, Germany). For recording, 5 μ l of an internal-like solution was applied to the internal side of the microchip. The internal solution contained: 50 mM CsCl, 10 mM NaCl, 2 mM MgCl₂, 60 mM CsF, 20 mM EGTA, and 10 mM HEPES, pH 7.2 and osmolarity 288 mOsM. Cs in the internal solution blocks potassium channel activity. A single cell suspension was added to an external solution of the following composition: 140 mM NaCl, 4 mM KCl, 1 mM MgCl₂, 2 mM CaCl₂F, 5 mM D-glucose monohydrate, and 10 mM HEPES, pH 7.4 and osmolarity 298 mOsM. Whole-cell currents were recorded using a ramp protocol ranging between -60 to $+130$ mV for 500 milliseconds. The mean membrane capacitance of N41 cells was 9 pF \pm 1 pF ($n = 10$). Mean access resistance was 15 ± 1 M Ω ($n = 10$). The holding potential (HP) was set to 0 mV in order to eliminate any possible contribution of VDCCs or sodium channels. All plots were generated with SigmaPlot software version 12.5 (Systat, San Jose, California, USA).

Data Evaluation and Statistics

GraphPad Prism 6.0 (GraphPad software, San Diego, Calif., USA) was chosen for visualization and data analysis. Data are shown as means \pm SEM of independent experiments. Statistical analysis was carried out using one-way and two-way ANOVA, followed by Sidak correction. For Ca²⁺ imaging, statistical significance was determined by an unpaired *t*-test with Welch's correction. In figure legends, the number of experiments and the type of comparison are given. Statistical significance was defined as * $p \leq 0.05$, ** $p \leq 0.01$, *** $p \leq 0.001$, and **** $p \leq 0.0001$.

RESULTS

Intraperitoneal Injection of 3-T₁AM Results in the Activation of PVN Neurons in C57BL/6 Mice

Previous studies demonstrated the accumulation and stimulatory effects of 3-T₁AM in selected brain nuclei such as locus coeruleus and PVN of the hypothalamus (6, 7). However, the exact role of this thyroid hormone metabolite in the hypothalamus remains unclear. To investigate whether 3-T₁AM is capable of activating hypothalamic neurons in these nuclei *in vivo*, we performed intraperitoneal injections of 50 mg/kg bodyweight of 3-T₁AM or DMSO/PBS as control, six mice per group, and monitored 3-T₁AM-induced neuron activation relative to DMSO/PBS. We used c-FOS as the marker for neuronal activity. One hour after 3-T₁AM injection, increased c-FOS staining of distinct neurons was clearly visible in the PVN (60 ± 13 c-FOS positive cells per brain slide), while DMSO/PBS treated mice showed only few c-FOS positive neurons (16 ± 4 c-FOS positive cells per brain slide) (Figures 1A,B). 3-T₁AM had no stimulatory effect on the medial preoptic area (MPO), the supraoptic nucleus (SON), the dorsolateral nucleus of the hypothalamus, the periaqueductal gray (PAG) and the ventral tegmental segment (VTA) (Figure 1B and Supplemental Figure 1). The c-FOS positive cells ranged from 4 to 25 per brain slide in these brain loci.

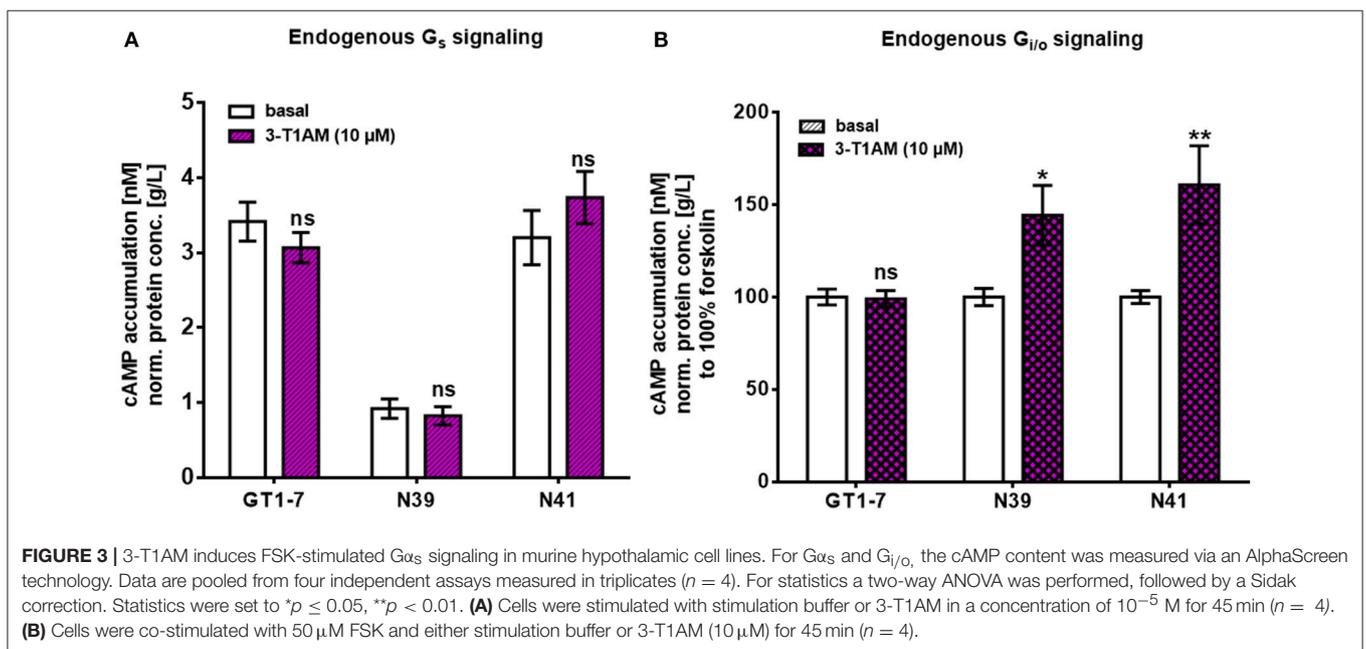
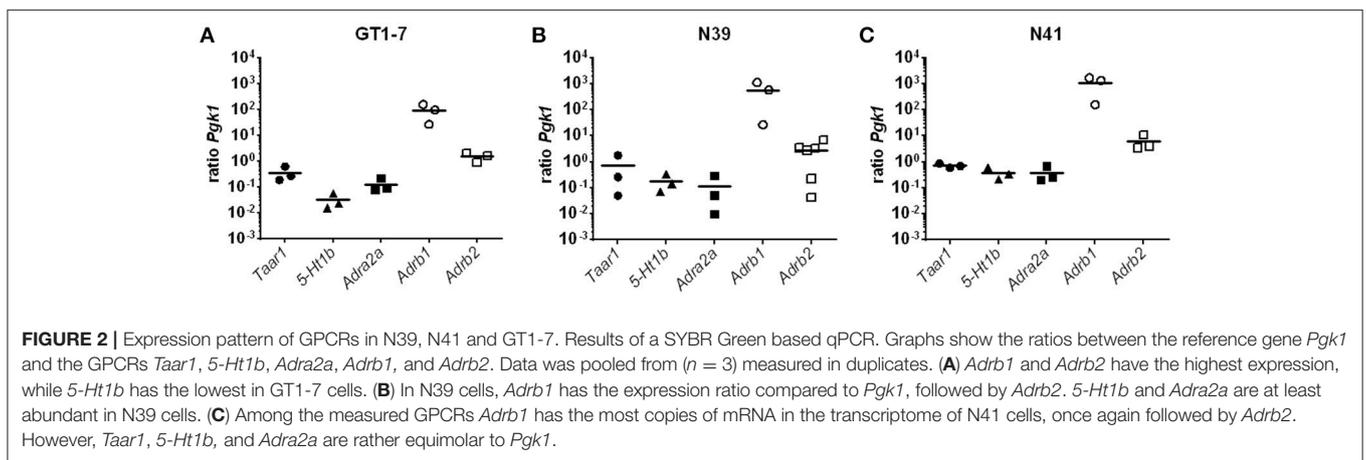
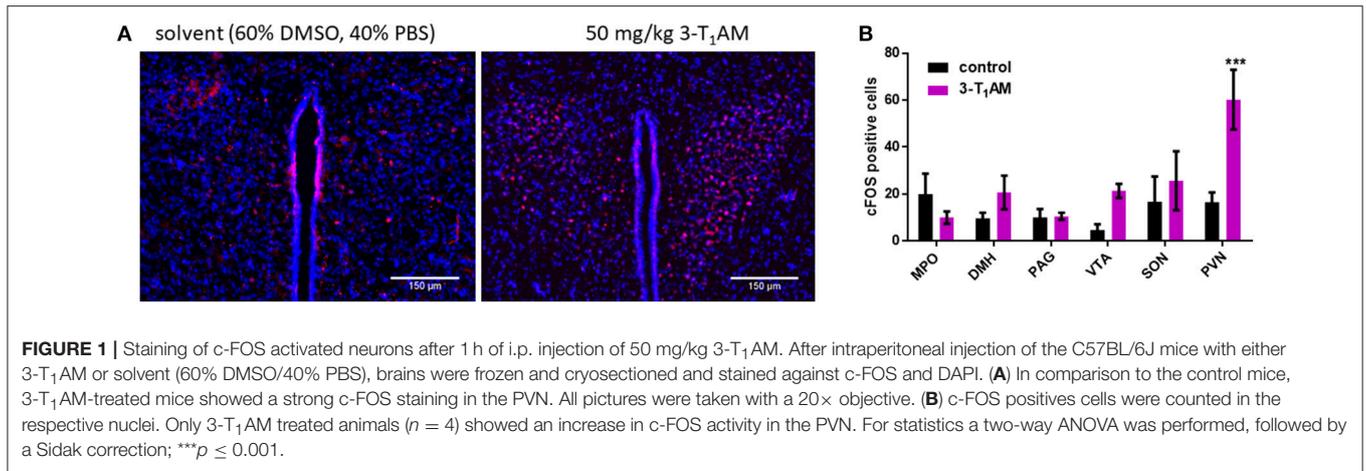
Expression Profile of GPCRs, the Main 3-T₁AM Target, in the Murine Hypothalamic Cell Lines

To elucidate the underlying mechanism behind the stimulatory effect of 3-T₁AM in the hypothalamus, we used three murine hypothalamic cell lines, GT1-7, mHypoE-N39 (N39), and mHypoE-N41 (N41). These cell lines are established models to study neuroendocrine mechanisms and known to express PVN-like markers (38, 42, 43). Here we demonstrated that in N41 cells, 3-T₁AM significantly increased the c-FOS activation (Supplemental Figure 2). As GPCRs are the primary targets of 3-T₁AM, we investigated the GPCR expression profile of the three cell lines. Here, we showed the expression of the aminergic receptors *Taar1*, *5-Ht1b*, *Adra2a*, *Adrb1*, and *Adrb2* (Figure 2). Furthermore, when compared against *Pgk1*, *Adrb1* noticeably had the highest expression rate among these receptors in all three cell lines with a ratio of 94 ± 38 for GT1-7, 565.6 ± 312 for N39, and 1046 ± 456 for N41. The second highest expression was detected for *Adrb2* with a ratio of 1.54 ± 0.34 for GT1-7, 2.77 ± 1.04 for N39, and 6.06 ± 2.34 for N41. The other three receptors *Taar1*, *5-Ht1b*, and *Adra2a*, displayed similar expression profiles. The mRNA content in all three cell lines was lower than the reference gene *Pgk1* (Figure 2), with ratios between 0.37 ± 0.14 for *5-Ht1b* in GT1-7 and up to 0.72 ± 0.08 for *Taar1* in N41.

3-T₁AM Induces FSK-Amplified G α_s Signaling in the Murine Hypothalamic Cell Lines

To investigate which pathway could contribute to 3-T₁AM actions in the hypothalamus, we tested two major signaling cascades downstream of 3-T₁AM GPCR targets, G α_s and G $\beta\gamma$. To measure endogenous G α_s signaling, we determined the cAMP enhancement. N41 and GT1-7 cells had a higher basal cAMP content with 3.2 ± 0.36 nM cAMP/g/L protein for N41 and 3.41 ± 0.26 nM cAMP/g/L protein for GT1-7, compared to N39 with 0.92 ± 0.13 nM cAMP/g/L protein (Figure 3A, $n = 4$ in triplicates, $p < 0.001$). In all cell lines, 3-T₁AM stimulation (10 μ M) did not increase cAMP concentration compared to the basal cAMP content (Figure 3A). Only NorEpi and ISOP activated an endogenous G α_s signal in GT1-7 (~ 1.6 fold for NorEpi and ~ 1.7 fold for ISOP), N41 (~ 1.8 fold for NorEpi and ~ 2.1 fold for ISOP), and N39 (~ 3.5 fold for NorEpi and ~ 5.9 fold for ISOP) cells (Supplemental Figure 3). 5-HT and PEA, endogenous ligands for 5-HT1b and TAAR1, did not increase cAMP content (Supplemental Figure 3).

To determine G $\beta\gamma$ signaling, cells were incubated with FSK, an unspecific activator of the adenylyl cyclase, which increases the cellular cAMP content. It is known that activation of G $\beta\gamma$ leads to inhibition of the adenylyl cyclase and a decrease in FSK-induced cAMP content. For all three cell lines, G $\beta\gamma$ activation was not detected after stimulation with 10 μ M 3-T₁AM (Figure 3B). In addition, FSK can potentiate weak G α_s signaling. Dessauer et al. showed that FSK in the presence of G α_s has a higher affinity to the adenylyl cyclase, yielding in a higher cAMP accumulation (44). Here, this phenomenon



emerges for N39 and N41 and 3-T₁AM stimulation significantly increases cAMP content in FSK-treated cells ($144.2 \pm 16.15\%$ for N39 and $160.77 \pm 21.17\%$ for N41, $n = 4$ in triplicates, $p_{N39} = 0.0128$, $p_{N41} = 0.0019$). The specific ligands for TAAR1, 5-HT1b, ADRA2A, ADRB1, and ADRB2 did not activate G_{i/o} signaling (Supplemental Figure 4). Collectively, in addition to an FSK-potentiated G α_s activation, 3-T₁AM had no detectable influence on cAMP content in the hypothalamic cell lines.

Gene Expression Profile of TRPs in Murine Hypothalamic Cell Lines

The aforementioned results demonstrated that GPCR-dependent signaling is not the sole regulator of 3-T₁AM-induced effects on hypothalamic cells. Previous studies identified several members of TRPs as new targets for 3-T₁AM (21–23). Here, we measured the gene expression levels of the TRPM subfamily and TRPV1 in three hypothalamic cell lines. The qPCR data show that none of the hypothalamic cell lines expressed *Trpm5* (Figure 4). Contrary to this, in GT1-7 cells, *Trpm4* was the highest expressed TRP channel with a ratio of 159 ± 89 compared to *Pgk1*, followed by *Trpm7* with a ratio of 11.7 ± 10.1 to *Pgk1*. *Trpv1* had a ratio of 2.96 ± 2.91 and *Trpm8* of 0.03 ± 0.02 to *Pgk1*. *Trpm1* was expressed with a ratio of 0.00075 ± 0.00045 to *Pgk1*. *Trpm6* had the lowest expression with a ratio of $0.00000064 \pm 0.00000031$ compared to *Pgk1* (Figure 4A). In N39 cells, *Trpm4* was also the highest expressed TRP channel with a ratio of 831 ± 617 to *Pgk1*, followed by *Trpv1* with a ratio of 79 ± 38 to *Pgk1*. *Trpm7* had a ratio of 59 ± 29 and *Trpm8* a ratio of 2.19 ± 0.72 . *Trpm1*, *Trpm2*, and *Trpm3* expression ratios laid between a ratio of 0.029 ± 0.025 to *Pgk1* for *Trpm2* and *Trpm3* with a ratio of 0.000089 ± 0.000076 to *Pgk1*. The mRNA content was comparable to GT1-7 cells, with *Trpm6* having the lowest expression with a ratio of 4.95 ± 3.48 compared to the reference gene (Figure 4B). N41 cells exhibited a similar expression pattern of TRP channels as GT1-7 and N39 cell lines. *Trpm4* (ratio to *Pgk1* 759 ± 674) and *Trpv1* (ratio to *Pgk1* 514 ± 226) were the highest expressed genes, followed by *Trpm7* (ratio to *Pgk1* 29 ± 23) and *Trpm8* (24 ± 18). *Trpm2* and *Trpm1* were lower expressed with ratios of 0.029 ± 0.025 and 0.0014 ± 0.0011 compared to the reference gene. *Trpm3* and *Trpm6* were least expressed in the transcriptome of N41 cells with ratios to *Pgk1* of 0.000089 ± 0.000076 and 0.0000022 ± 0.0000013 (Figure 4C).

3-T₁AM Increases Intracellular Ca²⁺ Concentration and Whole-Cell Currents in Murine Hypothalamic Cell Lines

To investigate the involvement of TRPs in the 3-T₁AM stimulatory effects in Ca²⁺ regulation, we monitored time-dependent changes in intracellular free Ca²⁺ levels ([Ca²⁺]_i) in single-cells. 3-T₁AM (10 μ M) increased the f_{340nm}/f_{380nm} ratio from 0.70 ± 0.01 to 0.77 ± 0.05 ; ($n = 10$) in GT1-7, from 0.70 ± 0.008 to 0.84 ± 0.03 ($n = 15$) in N39 cells and from 0.70 ± 0.009 to 1.88 ± 0.03 ; ($n = 15$; *** $p < 0.001$) in N41 cells (Figure 5). In untreated controls, this ratio remained constant at 0.70 ± 0.01 in GT1-7 cells ($n = 10$), 0.70 ± 0.009 in N39 cells ($n = 10$)

and 0.70 ± 0.01 in N41 cells after the same period ($n = 15$) (Figure 5). It should be noted that the strongest increase of Ca²⁺ concentration was detected in the N41 cells ($p \leq 0.0001$) which also has the highest expression level of adrenergic receptors and TRPs.

In the next step, we evaluated 3-T₁AM effects on whole-cell currents of N41 cells to determine if increases in their magnitude underlie rises in plasma membrane Ca²⁺ influx in this cell line. At -60 mV, 10 μ M 3-T₁AM increased inward currents from -14.38 pA/pF to -60.78 , which are attributable to Ca²⁺ influx because of the internal Ca²⁺ free solution. At $+130$ mV, outward rectifying currents strongly increased from 83.71 to 177.38 pA/pF in the presence of 3-T₁AM.

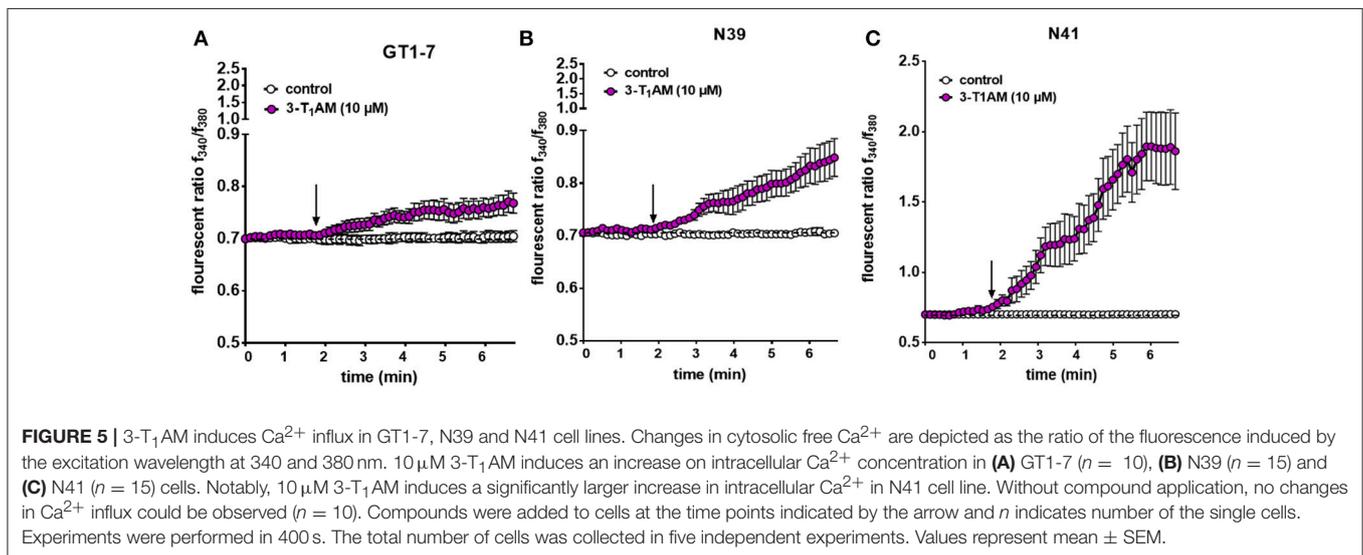
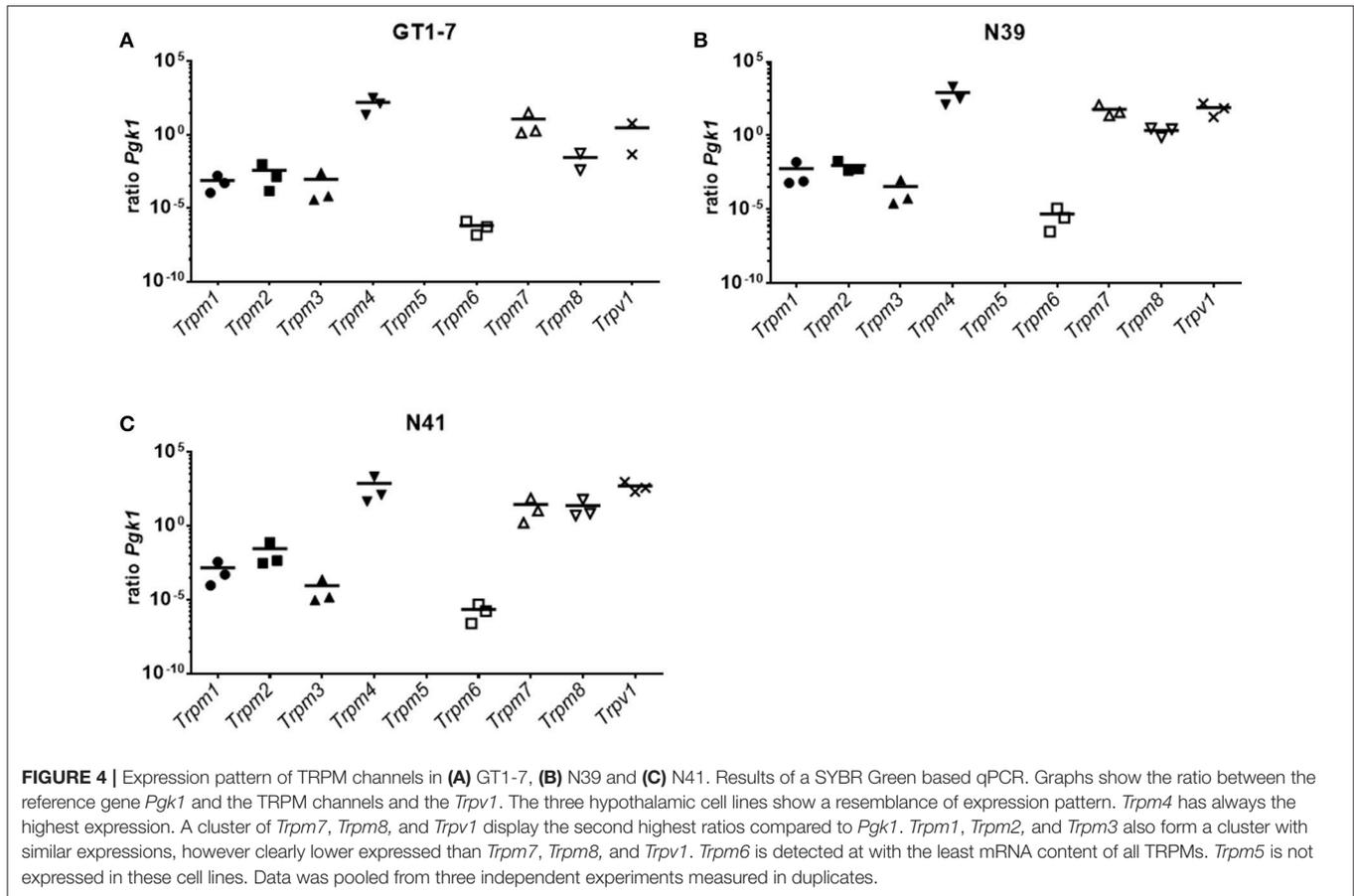
3-T₁AM Mediates Rises in Ca²⁺ Influx and Whole-Cell Currents Through TRPM8 Activation

Previous studies demonstrated that 3-T₁AM affects TRPM8 activation at a constant temperature in different cell types (21, 23). To validate that the Ca²⁺ increase stems from an increase in TRPM8 channel activity, N41 cells, which had the maximum response to 3-T₁AM stimulation, were exposed for 30 min to 10 μ M BCTC, followed by bath supplementation with 10 μ M 3-T₁AM. Under these conditions, the TRPM8 channel blocker abolished a 3-T₁AM-induced Ca²⁺ rise. More specifically, the f_{340nm}/f_{380nm} ratio decreased from 1.81 ± 0.04 to 0.93 ± 0.02 in the presence of BCTC ($n = 15$) ($p \leq 0.001$) (Figure 6A and Supplemental Figure 5).

Our previous study demonstrated the inverse association between TRPM8 and TRPV1 induced by 3-T₁AM (21, 22). Different observations showed BCTC acts as a non-specific TRPV1 inhibitor (45, 46). Here, we also demonstrated the high gene expression of TRPV1 in murine hypothalamic cell lines. To rule out the involvement of TRPV1 in the 3-T₁AM-induced intracellular Ca²⁺ response, we used AMTB as a high selective TRPM8 blocker and capsazepine (CPZ) as a specific TRPV1 blocker. In the presence of 10 μ M AMTB, the f_{340nm}/f_{380nm} ratio decreased from 1.83 ± 0.08 to 1.12 ± 0.05 (Figure 6B), whereas 10 μ M CPZ had no significant inhibitory effect on 3-T₁AM-induced intracellular Ca²⁺ response (Supplemental Figure 6). As AMTB suppressed a 3-T₁AM-induced Ca²⁺ increase, we validated this effect by determining if this inhibitor influenced underlying whole-cell currents. In the presence of 10 μ M AMTB, inward currents decreased to -14.61 pA/pF and outward currents decreased to 82.40 pA/pF (Figure 7). Considering all these findings, 3-T₁AM increased intracellular Ca²⁺ concentration and whole-cell currents in mouse hypothalamic cells, thus confirming similar effects in other cell types.

DISCUSSION

Administration of 3-T₁AM in mice results in reversible effects such as reduction of body temperature, cardiac output, and the respiratory quotient along with anapyrexia and hyperglycemia (1, 47). There is evidence that 3-T₁AM accumulates in the



hypothalamic nuclei (6, 7). The aim of this study was to explore the underlying mechanism behind the stimulatory effect of this thyroid hormone metabolite in selected hypothalamic regions.

3-T₁AM-Induced Signaling Network Activates PVN Neurons of C57BL/6 Mice

Within the hypothalamus, PVN is one of the most extensively studied nuclei and is playing a pivotal role in the control of

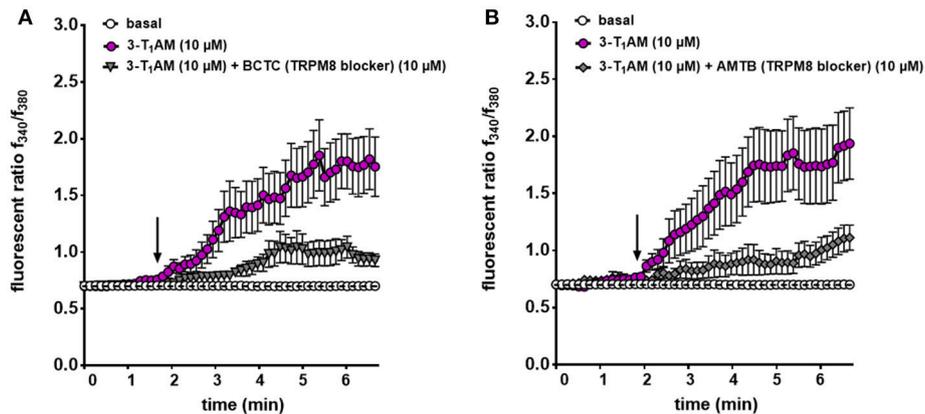


FIGURE 6 | TRPM8 mediates 3-T₁AM-induced Ca²⁺ response in N41 cell line. Cells were pre-incubated with inhibitors (10 μM AMTB or 10 μM BCTC) 30 min before the measurement. Stimulation was performed with 10 μM 3-T₁AM and Ca²⁺ influxes were measured ($n = 15-19$) with and without the inhibitors. **(A)** 3-T₁AM increased Ca²⁺ influx and pre-incubation with BCTC significantly suppressed this effect. **(B)** AMTB showed the similar inhibitory effect on 3-T₁AM-induced Ca²⁺ influx. Experiments were performed in 400 s. Compounds were added to cells at the time points indicated by the arrow and n indicates number of the single cells. The total number of cells was collected in 5 independent experiments. Values represent mean \pm SEM.

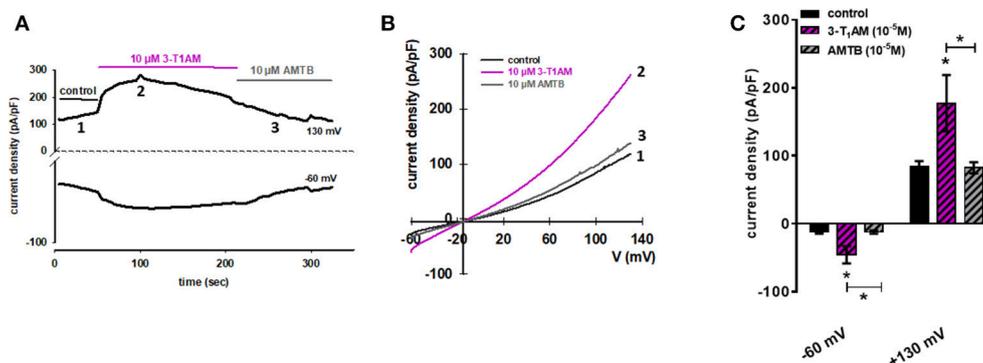


FIGURE 7 | 3-T₁AM activates whole-cell currents in N41 cell line. **(A)** Time course of whole-cell currents at -60 mV (lower trace) and 130 mV (upper trace) showing the current activation by $10 \mu\text{M}$ 3-T₁AM (left). **(B)** Original traces of 3-T₁AM activated current responses to voltage ramps from -60 mV up to $+130$ mV (right). Currents are shown before application (labeled as 1) and during application of $1 \mu\text{M}$ 3-T₁AM (labeled as 2) and in the presence of $10 \mu\text{M}$ AMTB (labeled as 3). **(C)** $10 \mu\text{M}$ 3-T₁AM increased inward and outward whole-cell channel currents in N41 cells and this effect was strongly suppressed in the presence of $10 \mu\text{M}$ AMTB. Currents are shown before application (control), during application of $10 \mu\text{M}$ 3-T₁AM and in the presence of $10 \mu\text{M}$ AMTB ($n = 10$). Whole-cell currents were recorded using step and ramp protocols involving voltage steps of 10 mV ranging between -60 to $+130$ mV for 400 ms. The currents were normalized to capacitance to obtain current density (pA/pF). Statistical significance was determined by one-way ANOVA, comparing the basal current density (pA/pF) against 3-T₁AM and AMTB. Data are the mean \pm SEM of at least 10 independent experiments; $*p \leq 0.05$.

fluid homeostasis, lactation, cardiovascular regulation, feeding behavior, nociception and response to stress (11). It has already been shown that the direct injection of 3-T₁AM into the lateral ventricle of male mice leads to the activation of neurons in the anterior commissural nucleus of hypothalamus (48). Here, we observed that intraperitoneal injection of 3-T₁AM results in the activation of PVN neurons in C57BL/6 mice (Figure 1). Recently, Gachkar et al. showed that 3-T₁AM-induced hypothermia is due to vasodilation, which is not directly induced in the veins. In this study they suggested that 3-T₁AM might induce the tail vasodilation through central action in male mice (14). Here, we investigated the loci in the brain, which are known to be involved in the

regulation of the body temperature through non-shivering and shivering thermogenesis (PAG and DMH) and heat dissipation like vasodilation [VTA, (49)]. The result of this study demonstrated no differences in c-FOS activation in these areas (Figure 1B).

Previous studies detected no effect of 3-T₁AM metabolites such as 3-iodothyroacetic acid and iodine-free thyronamine on cardiac output or thermoregulation (50, 51). Nonetheless, whether 3-T₁AM directly activates the PVN neurons or causes the activation of neuronal projections from other nuclei is uncertain. Moreover, the exact cellular mechanism initiated by 3-T₁AM once it reaches in the hypothalamic nuclei is yet to be discovered.

3-T₁AM Slightly Stimulates G α_s Signaling in Murine Hypothalamic Cell Lines

Aminergic receptors are largely expressed in the hypothalamus and play a substantial role in various regulatory responses, such as metabolic regulations (52). These receptors are known 3-T₁AM targets and previous studies assumed that pharmacological effects of this thyroid hormone metabolite are attributable to aminergic receptor signaling (1, 16–19). It has been reported that 3-T₁AM enhances G α_s signaling in response to ISOP co-stimulation at ADRB2 (17) and activates G i/o signaling combined with NorEpi of the ADRA2A in transfected HEK293 cells (16). Here, we could not detect the activation of G i/o in response to the 3-T₁AM in murine hypothalamic cell lines (**Figure 3B**). Nevertheless, N39 and N41 showed an FSK-amplified G α_s signal due to 3-T₁AM stimulation (**Figure 3B**). The low GPCR expression rate could be the reason for the lack of strong G α_s in these cells (**Figure 2**). The absence of G i/o signal through 5-HT1b could be explained by dimerization with TAAR1 (19). It was shown that 3-T₁AM is capable of inducing a clear G i/o signal in HEK293 cells overexpressing 5-HT1b. However, co-expression of 5-HT1b with TAAR1 abrogated 3-T₁AM-induced cAMP signaling (19). Since the three hypothalamic cell lines all co-express TAAR1 and 5-HT1b, the G i/o signal of 5-HT1b via 3-T₁AM stimulation is probably disrupted. The FSK stimulated N39 and N41 cells showed a significant increase in cAMP concentration after 3-T₁AM challenge (**Figure 3B**). Besides its own adenylyl cyclase activating property, FSK stimulation additionally favors activation of adenylyl cyclase through G α_s (44, 53, 54). Here, we concluded that 3-T₁AM induces a weak G α_s signal in N39 and N41 cells.

3-T₁AM Activates TRPM8 Channel in Murine Hypothalamic Cell Lines

Ca²⁺ mobilization in different cell types plays an essential role in the modulation of c-FOS gene expression (55, 56). In neurons, Ca²⁺ influx is a prerequisite for activation of the ERK/MAPK pathway. It is known that c-FOS activation reflects summation or integration of this Ca²⁺ dependent-neuronal activity (57, 58). Moreover, Ca²⁺ acts synergistically with cAMP to activate c-FOS transcription (59). To test whether 3-T₁AM induces an increase of intracellular Ca²⁺ in hypothalamic cell lines, we evaluated ion channel activation induced by 3-T₁AM.

It is known that GPCR activation induces an increase of intracellular Ca²⁺ concentration through different pathways (60–63). Recent studies demonstrated the co-expression of GPCRs and TRPs in a variety of cell types (31). Different signaling intermediates such as adaptor proteins, kinases, and lipid metabolites functionally link GPCRs to TRPs (30, 64).

Various TRPs play a pivotal role in the mechanisms that are involved in energy metabolism and temperature adaptation (26, 65, 66). Some recent studies reported that activation of warm-sensitive TRPM2 leads to a similar thermoregulatory response as the one observed in mice after systemic administration of 3-T₁AM (67). Previously, we showed that 3-T₁AM acts as a cooling agent to directly affect TRPM8 activation in different cell types (21, 22). In rat thyrocyte, 3-T₁AM induced

Ca²⁺ responses similar to specific TRPM8 agonists such as menthol (68). In ocular cells, 3-T₁AM evoked Ca²⁺ mobilization and increases in whole-cell currents, a stimulatory effect that could be specifically attenuated in the presence of specific TRPM8 blocker (21, 22). The result of this study showed that 3-T₁AM induces intracellular Ca²⁺ increase through the TRPM8 channel in murine hypothalamic cell lines. Finding the functional link between 3-T₁AM and this specific TRP channel in hypothalamic cell lines is relevant since there are indications that the TRPM8 channel regulates energy metabolism in different tissues and plays a crucial role in thermoregulation (69, 70). TRPM8 stimulation, for instance, induces mitochondrial activation and heat production in brown adipocytes. Chronic TRPM8 agonist administration enhances the energy metabolism in brown adipocytes and prevents obesity in mice (69). In skeletal muscles, TRPM8 activation by dietary menthol improves energy metabolism through Ca²⁺-dependent upregulation of the peroxisome proliferator-activated receptor- γ coactivator 1 α (PGC1 α) which is involved in the mitochondrial function (70). Recently, it has been demonstrated that TRPM8-deficient mice develop late-onset obesity and metabolic dysfunction at moderate cooling, suggesting the importance of TRPM8 in the coupling between thermoregulation and energy homeostasis. Nevertheless, *Trpm8*^{-/-} mice exhibit a remarkable decrease of core body temperature due to increased tail heat loss (71) which is in contrast to the 3-T₁AM stimulatory effect on TRPM8 found in our *in vitro* electrophysiological observations. Therefore, we investigated the possible effect of 3-T₁AM on TRPV1 as another known thermo-sensitive TRP channel as it has been shown that there is an interplay between TRPV1 and TRPM8 (72).

Previous studies demonstrated the co-expression of TRPV1 with TRPM8 in different cell types including rat hippocampal neurons, intralobar pulmonary arteries, aorta, neuroendocrine tumor cells, retinoblastoma cells, uveal melanoma cells, and corneal cells (36, 73–76). Here, we showed the co-expression of these two thermo-TRP channels in three different hypothalamic cell lines. It is well established that there is a cross talk between TRPM8 and TRPV1 channels in various tissues. For instance, TRPM8 agonist blocks the mechanical and heat hyperalgesia caused by TRPV1 activation (77, 78). It was also demonstrated that icilin, a specific TRPM8 agonist, attenuates TRPV1-dependent calcitonin gene-related peptide release in the colon and is suggested as a promising therapeutic target for the treatment of colitis (79). The interdependence of the TRPM8 and TRPV1 ion channel function as well as the role of both channels in thermo-regulation have raised the question as to which TRP channel is the main target of 3-T₁AM in hypothalamic cell lines. The results of this study clearly showed that the effect of 3-T₁AM were not attributable to TRPV1 since only the specific TRPM8 blocker (AMTB) could strongly inhibit the 3-T₁AM-induced Ca²⁺ influx and whole-cell current.

Collectively, the results of this study demonstrated the Ca²⁺ signal transduction pathways induced by 3-T₁AM and provided evidence of TRP channel modulation via this TH derivative in hypothalamic cell lines. Characterization of the intracellular signaling cascade induced by 3-T₁AM might explain

the underlying mechanism behind the profound physiological effects of this metabolite.

AUTHOR CONTRIBUTIONS

HB, NK, and JB designed the study. JB, SJ, and NK performed the experiments and analyzed the data. SJ, CH, and JM performed the mouse studies. NK, JB, SM, and HB wrote and edited the manuscript. CH, JM, and MR discussed data and edited the manuscript. All authors approved the manuscript.

FUNDING

This work was supported by the Deutsche Forschungsgemeinschaft (DFG), the priority program SPP1629

REFERENCES

- Scanlan TS, Suchland KL, Hart ME, Chiellini G, Huang Y, Kruzich PJ, et al. 3-iodothyronamine is an endogenous and rapid-acting derivative of thyroid hormone. *Nat Med.* (2004) 10:638–42. doi: 10.1038/nm1051
- Hoefig CS, Zucchi R, Kohrle J. Thyronamines and derivatives: physiological relevance, pharmacological actions, and future research directions. *Thyroid* (2016) 26:1656–73. doi: 10.1089/thy.2016.0178
- Hoefig CS, Köhrle J, Brabant G, Dixit K, Yap B, Strasburger CJ, et al. Evidence for extrathyroidal formation of 3-iodothyronamine in humans as provided by a novel monoclonal antibody-based chemiluminescent serum immunoassay. *J Clin Endocrinol Metab.* (2011) 96:1864–72. doi: 10.1210/jc.2010-2680
- Galli E, Marchini M, Saba A, Berti S, Tonacchera M, Vitti P, et al. Detection of 3-iodothyronamine in human patients: a preliminary study. *J Clin Endocrinol Metab.* (2012) 97:E69–74. doi: 10.1210/jc.2011-1115
- Klieverik LP, Foppen E, Ackermans MT, Serlie MJ, Sauerwein HP, Scanlan TS, et al. Central effects of thyronamines on glucose metabolism in rats. *J Endocrinol.* (2009) 201:377–86. doi: 10.1677/JOE-09-0043
- Chiellini G, Erba P, Carnicelli V, Manfredi C, Frascarelli S, Ghelardoni S, et al. Distribution of exogenous [125I]-3-iodothyronamine in mouse *in vivo*: relationship with trace amine-associated receptors. *J Endocrinol.* (2012) 213:223–30. doi: 10.1530/JOE-12-0055
- Gompf HS, Greenberg JH, Aston-Jones G, Ianculescu AG, Scanlan TS, Dratman MB. 3-Monoiodothyronamine: the rationale for its action as an endogenous adrenergic-blocking neuromodulator. *Brain Res.* (2010) 1351:130–40. doi: 10.1016/j.brainres.2010.06.067
- Fliers E, Alkemade A, Wiersinga WM, Swaab DF. Hypothalamic thyroid hormone feedback in health and disease. *Prog Brain Res.* (2006) 153:189–207. doi: 10.1016/S0079-6123(06)53011-0
- Lukoyanov NV, Lukoyanova EA. Retrosplenial cortex lesions impair acquisition of active avoidance while sparing fear-based emotional memory. *Behav Brain Res.* (2006) 173:229–36. doi: 10.1016/j.bbr.2006.06.026
- Timmann D, Daum I. Cerebellar contributions to cognitive functions: a progress report after two decades of research. *Cerebellum* (2007) 6:159. doi: 10.1080/14734220701496448
- Ferguson AV, Latchford KJ, Samson WK. The paraventricular nucleus of the hypothalamus A potential target for integrative treatment of autonomic dysfunction. *Expert Opin Ther Targets* (2008) 12:717–27. doi: 10.1517/14728222.12.6.717
- Manni ME, De Siena G, Saba A, Marchini M, Dicembrini I, Bigagli E, et al. 3-Iodothyronamine: a modulator of the hypothalamus-pancreas-thyroid axes in mice. *Br J Pharmacol.* (2012) 166:650–8. doi: 10.1111/j.1476-5381.2011.01823.x
- Manni ME, De Siena G, Saba A, Marchini M, Landucci E, Gerace E, et al. Pharmacological effects of 3-iodothyronamine (T1AM) in mice include facilitation of memory acquisition and retention and

reduction of pain threshold. *Br J Pharmacol.* (2013) 168:354–62. doi: 10.1111/j.1476-5381.2012.02137.x

ACKNOWLEDGMENTS

The authors would like to thank Cigdem Cetindag (Charité, Institute of Experimental Pediatric Endocrinology) for the technical assistance.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fendo.2018.00523/full#supplementary-material>

- Gachkar S, Oelkrug R, Martinez-Sanchez N, Rial-Pensado E, Warner A, Hoefig CS, et al. 3-iodothyronamine induces tail vasodilation through central action in male mice. *Endocrinology* (2017) 158:1977–84. doi: 10.1210/en.2016-1951
- Kleinau G, Pratzka J, Nurnberg D, Gruters A, Fuhrer-Sakel D, Krude H, et al. (2011). Differential modulation of Beta-adrenergic receptor signaling by trace amine-associated receptor 1 agonists. *PLoS ONE* 6:e27073. doi: 10.1371/journal.pone.0027073
- Dinter J, Mühlhaus J, Jacobi SE, Wienchol CL, Cöster M, Meister J, et al. 3-iodothyronamine differentially modulates α -2A-adrenergic receptor-mediated signaling. *J Mol Endocrinol.* (2015) 54:205–16. doi: 10.1530/JME-15-0003
- Dinter J, Khajavi N, Mühlhaus J, Wienchol CL, Cöster M, Hermsdorf T, et al. The multitarget ligand 3-iodothyronamine modulates β -adrenergic receptor 2 signaling. *Eur Thyroid J.* (2015) 4:21–9. doi: 10.1159/000381801
- Laurino A, Matucci R, Vistoli G, Raimondi L. 3-iodothyronamine (T1AM), a novel antagonist of muscarinic receptors. *Eur J Pharmacol.* (2016) 793:35–42. doi: 10.1016/j.ejphar.2016.10.027
- Bräunig J, Dinter J, Höfig CS, Paisdzior S, Szczepek M, Scheerer P, et al. The trace amine-associated receptor 1 agonist 3-iodothyronamine induces biased signaling at the serotonin 1b receptor. *Front Pharmacol.* (2018) 9:222. doi: 10.3389/fphar.2018.00222
- Ghelardoni S, Suffredini S, Frascarelli S, Brogioni S, Chiellini G, Ronca-Testoni S, et al. Modulation of cardiac ionic homeostasis by 3-iodothyronamine. *J Cell Mol Med.* (2009) 13:3082–90. doi: 10.1111/j.1582-4934.2009.00728.x
- Khajavi N, Reinach PS, Slavi N, Skrzypski M, Lucius A, Strauss O, et al. Thyronamine induces TRPM8 channel activation in human conjunctival epithelial cells. *Cell Signal* (2015) 27:315–25. doi: 10.1016/j.cellsig.2014.11.015
- Lucius A, Khajavi N, Reinach PS, Kohrle J, Dhandapani P, Huimann P, et al. 3-Iodothyronamine increases transient receptor potential melastatin channel 8 (TRPM8) activity in immortalized human corneal epithelial cells. *Cell Signal* (2016) 28:136–47. doi: 10.1016/j.cellsig.2015.12.005
- Khajavi N, Mergler S, Biebermann H. 3-iodothyronamine, a novel endogenous modulator of transient receptor potential melastatin 8? *Front Endocrinol.* (2017) 8:198. doi: 10.3389/fendo.2017.00198
- Liu BY, Fan L, Balakrishna S, Sui AW, Morris JB, Jordt SE. TRPM8 is the principal mediator of menthol-induced analgesia of acute and inflammatory pain. *Pain* (2013) 154:2169–77. doi: 10.1016/j.pain.2013.06.043
- Pingle S, Matta J, Ahern G. Capsaicin receptor: TRPV1 a promiscuous TRP channel. In: Flockerzi V, Nilius B, editors. *Transient Receptor Potential (TRP) Channels*. Berlin; Heidelberg: Springer (2007) 155–71.
- Voronova IP, Tuzhikova AA, Kozyreva TV. (2014). Expression of Genes for Temperature-Sensitive TRP Channels in the Rat Hypothalamus in Normal

- Conditions and on Adaptation to Cold. *Neurosci Behav Physiol.* 44:565–70. doi: 10.1007/s11055-014-9952-z
27. Axelsson HE, Minde JK, Sonesson A, Toolanen G, Hogestatt ED, Zygmunt PM. Transient receptor potential vanilloid 1, vanilloid 2 and melastatin 8 immunoreactive nerve fibers in human skin from individuals with and without norrbottnian congenital insensitivity to pain. *Neuroscience* (2009) 162:1322–32. doi: 10.1016/j.neuroscience.2009.05.052
 28. Robbins A, Kurose M, Winterson BJ, Meng ID. Menthol activation of corneal cool cells induces TRPM8-mediated lacrimation but not nociceptive responses in rodents. *Invest Ophthalmol Vis Sci.* (2012) 53:7034–42. doi: 10.1167/iiov.12-10025
 29. Alamri A, Bron R, Brock JA, Ivanusic JJ. Transient receptor potential cation channel subfamily V member 1 expressing corneal sensory neurons can be subdivided into at least three subpopulations. *Front Neuroanat.* (2015) 9:71. doi: 10.3389/fnana.2015.00071
 30. Clapham DE. TRP channels as cellular sensors. *Nature* (2003) 426:517–24. doi: 10.1038/nature02196
 31. Veldhuis NA, Poole DP, Grace M, McIntyre P, Bunnett NW. The G protein-coupled receptor–transient receptor potential channel axis: molecular insights for targeting disorders of sensation and inflammation. *Pharmacol Rev.* (2015) 67:36–73. doi: 10.1124/pr.114.009555
 32. Kunert-Keil C, Bisping F, Kruger J, Brinkmeier H. Tissue-specific expression of TRP channel genes in the mouse and its variation in three different mouse strains. *BMC Genomics* (2006) 7:159. doi: 10.1186/1471-2164-7-159
 33. Qiu J, Fang Y, Ronnekleiv OK, Kelly MJ. Leptin excites proopiomelanocortin neurons via activation of TRPC channels. *J Neurosci.* (2010) 30:1560–5. doi: 10.1523/JNEUROSCI.4816-09.2010
 34. Moran MM, McAlexander MA, Biro T, Szallasi A. Transient receptor potential channels as therapeutic targets. *Nat Rev Drug Discov.* (2011) 10:601–20. doi: 10.1038/nrd3456
 35. Sukumar P, Sedo A, Li J, Wilson LA, O'regan D, Lippiat JD, et al. Constitutively active TRPC channels of adipocytes confer a mechanism for sensing dietary fatty acids and regulating adiponectin. *Circ Res.* (2012) 111:191–200. doi: 10.1161/CIRCRESAHA.112.270751
 36. Mergler S, Valtink M, Takayoshi S, Okada Y, Miyajima M, Saika S, et al. Temperature-sensitive transient receptor potential channels in corneal tissue layers and cells. *Ophthalmic Res.* (2014) 52:151–9. doi: 10.1159/000365334
 37. Reinach PS, Mergler S, Okada Y, Saika S. Ocular transient receptor potential channel function in health and disease. *BMC ophthalmol.* (2015) 15(Suppl. 1):153. doi: 10.1186/s12886-015-0135-7
 38. Mellon PL, Windle JJ, Goldsmith PC, Padula CA, Roberts JL, Weiner RI. Immortalization of hypothalamic GnRH neurons by genetically targeted tumorigenesis. *Neuron* (1990) 5:1–10. doi: 10.1016/0896-6273(90)90028-E
 39. Belsham DD, Cai F, Cui H, Smukler SR, Salapatek AMF, Shkreta L. Generation of a phenotypic array of hypothalamic neuronal cell models to study complex neuroendocrine disorders. *Endocrinology* (2004) 145:393–400. doi: 10.1210/en.2003-0946
 40. Boda E, Pini A, Hoxha E, Parolisi R, Tempia F. Selection of reference genes for quantitative real-time RT-PCR studies in mouse brain. *J Mol Neurosci.* (2009) 37:238–53. doi: 10.1007/s12031-008-9128-9
 41. Khajavi N, Reinach PS, Skrzypski M, Lude A, Mergler S. L-carnitine reduces in human conjunctival epithelial cells hypertonic-induced shrinkage through interacting with TRPV1 channels. *Cell Physiol Biochem.* (2014) 34:790–803. doi: 10.1159/000363043
 42. Wetsel WC, Mellon PL, Weiner RI, Negro-Vilar A. Metabolism of proluteinizing hormone-releasing hormone in immortalized hypothalamic neurons. *Endocrinology* (1991) 129:1584–95. doi: 10.1210/endo-129-3-1584
 43. Mayer CM, Fick LJ, Gingerich S, Belsham DD. Hypothalamic cell lines to investigate neuroendocrine control mechanisms. *Front Neuroendocrinol.* (2009) 30:405–23. doi: 10.1016/j.yfrne.2009.03.005
 44. Dessauer CW, Scully TT, Gilman AG. Interactions of forskolin and ATP with the cytosolic domains of mammalian adenylyl cyclase. *J Biol Chem.* (1997) 272:22272–7. doi: 10.1074/jbc.272.35.22272
 45. Yan L, Wang J, Pan M, Qiu Q, Huang W, Qian H. Synthesis of analogues of BCTC incorporating a pyrrolidinyll linker and biological evaluation as transient receptor potential vanilloid 1 antagonists. *Chem Biol Drug Des.* (2016) 87:306–11. doi: 10.1111/cbdd.12661
 46. Schwarz MG, Namer B, Reeh PW, Fischer MJM. TRPA1 and TRPV1 antagonists do not inhibit human acidosis-induced pain. *J Pain* (2017) 18:526–34. doi: 10.1016/j.jpain.2016.12.011
 47. Regard JB, Kataoka H, Cano DA, Camerer E, Yin L, Zheng YW, et al. Probing cell type-specific functions of Gi *in vivo* identifies GPCR regulators of insulin secretion. *J Clin Invest.* (2007) 117:4034–43. doi: 10.1172/JCI32994
 48. Dhillo WS, Bewick GA, White NE, Gardiner JV, Thompson EL, Bataveljic A, et al. The thyroid hormone derivative 3-iodothyronamine increases food intake in rodents. *Diabetes Obes Metab.* (2009) 11:251–60. doi: 10.1111/j.1463-1326.2008.00935.x
 49. Tan CL, Knight ZA. Regulation of body temperature by the nervous system. *Neuron* (2018) 98:31–48. doi: 10.1016/j.neuron.2018.02.022
 50. Hoefig CS, Jacobi SE, Warner A, Harder L, Schanze N, Vennstrom B, et al. 3-Iodothyroacetic acid lacks thermoregulatory and cardiovascular effects *in vivo*. *Br J Pharmacol.* (2015) 172:3426–33. doi: 10.1111/bph.13131
 51. Harder L, Schanze N, Sarsenbayeva A, Kugel F, Kohrle J, Schomburg L, et al. *In vivo* Effects of repeated thyronamine administration in male C57BL/6J Mice. *Eur Thyroid J.* (2018) 7:3–12. doi: 10.1159/000481856
 52. Hazell GGJ, Hindmarch CC, Pope GR, Roper JA, Lightman SL, Murphy D, et al. G protein-coupled receptors in the hypothalamic paraventricular and supraoptic nuclei – serpentine gateways to neuroendocrine homeostasis. *Front Neuroendocrinol.* (2012) 33:45–66. doi: 10.1016/j.yfrne.2011.07.002
 53. Daly JW, Padgett W, Seamon KB. Activation of cyclic AMP-generating systems in brain membranes and slices by the diterpene forskolin: augmentation of receptor-mediated responses. *J Neurochem.* (1982) 38:532–44. doi: 10.1111/j.1471-4159.1982.tb08660.x
 54. Darfler FJ, Mahan LC, Koachman AM, Insel PA. Stimulation of forskolin of intact S49 lymphoma cells involves the nucleotide regulatory protein of adenylyl cyclase. *J Biol Chem.* (1982) 257:11901–7.
 55. Ran W, Dean M, Levine RA, Henkle C, Campisi J. Induction of C-Fos and C-Myc messenger-Rna by epidermal growth-factor or calcium ionophore is camp dependent. *Proc Natl Acad Sci USA.* (1986) 83:8216–20. doi: 10.1073/pnas.83.21.8216
 56. Li SL, Godson C, Roche E, Zhao SJ, Prentki M, Schlegel W. Induction of c-fos in pituitary cells by thyrotrophin-releasing hormone and phorbol 12-myristate 13-acetate depends upon Ca2+ influx. *J Mol Endocrinol.* (1994) 13:303–12. doi: 10.1677/jme.0.0130303
 57. Deisseroth K, Mermelstein PG, Xia H, Tsien RW. Signaling from synapse to nucleus: the logic behind the mechanisms. *Curr Opin Neurobiol.* (2003) 13:354–65. doi: 10.1016/S0959-4388(03)00076-X
 58. Cohen S, Greenberg MF. Communication between the synapse and the nucleus in neuronal development, plasticity, and disease. *Annu Rev Cell Dev Biol.* (2008) 24:183–209. doi: 10.1146/annurev.cellbio.24.110707.175235
 59. Coulon V, Veyrune JL, Tourkine N, Vié A, Hipskind RA, Blanchard JM. A Novel Calcium signaling pathway targets the c-fosIntragenic transcriptional pausing site. *J Biol Chem.* (1999) 274:30439–46. doi: 10.1074/jbc.274.43.30439
 60. Barritt GJ. Receptor-activated Ca²⁺ inflow in animal cells: a variety of pathways tailored to meet different intracellular Ca²⁺ signalling requirements. *Biochem J.* (1999) 337(Pt 2):153–169.
 61. Chen Y, Geis C, Sommer C. Activation of TRPV1 contributes to morphine tolerance: involvement of the mitogen-activated protein kinase signaling pathway. *J Neurosci.* (2008) 28:5836–45. doi: 10.1523/JNEUROSCI.4170-07.2008
 62. Shen Y, Rampino MA, Carroll RC, Navy S. G-protein-mediated inhibition of the Trp channel TRPM1 requires the Gbetagamma dimer. *Proc Natl Acad Sci USA.* (2012) 109:8752–7. doi: 10.1073/pnas.1117433109
 63. Yekkirala AS. Two to tango: GPCR oligomers and GPCR-TRP channel interactions in nociception. *Life Sci.* (2013) 92:438–45. doi: 10.1016/j.lfs.2012.06.021
 64. Petho G, Reeh PW. Sensory and signaling mechanisms of bradykinin, eicosanoids, platelet-activating factor, and nitric oxide in peripheral nociceptors. *Physiol Rev.* (2012) 92:1699–775. doi: 10.1152/physrev.00048.2010
 65. Michlig S, Merlini JM, Beaumont M, Ledda M, Tavenard A, Mukherjee R, et al. Effects of TRP channel agonist ingestion on metabolism and autonomic nervous system in a randomized clinical trial of healthy subjects. *Sci Rep.* (2016) 6:20795. doi: 10.1038/srep20795

66. Wang XP, Yu X, Yan XJ, Lei F, Chai YS, Jiang JF, et al. TRPM8 in the negative regulation of TNF α expression during cold stress. *Sci Rep.* (2017) 7:45155. doi: 10.1038/srep45155
67. Song K, Wang H, Kamm GB, Pohle J, Reis FD, Heppenstall P, et al. The TRPM2 channel is a hypothalamic heat sensor that limits fever and can drive hypothermia. *Science* (2016) 353:1393–8. doi: 10.1126/science.aaf7537
68. Schanze N, Jacobi SE, Rijntjes E, Mergler S, Del Olmo M, Hoefig CS, et al. 3-iodothyronamine decreases expression of genes involved in iodide metabolism in mouse Thyroids and inhibits iodide uptake in PCCL3 thyrocytes. *Thyroid* (2017) 27:11–22. doi: 10.1089/thy.2016.0182
69. Ma S, Yu H, Zhao Z, Luo Z, Chen J, Ni Y, et al. Activation of the cold-sensing TRPM8 channel triggers UCP1-dependent thermogenesis and prevents obesity. *J Mol Cell Biol.* (2012) 4:88–96. doi: 10.1093/jmcb/mjs001
70. Li C, Li J, Xiong X, Liu Y, Lv Y, Qin S, et al. TRPM8 activation improves energy expenditure in skeletal muscle and exercise endurance in mice. *Gene* (2018) 641:111–6. doi: 10.1016/j.gene.2017.10.045
71. Reimundez A, Fernandez-Pena C, Garcia G, Fernandez R, Ordas P, Gallego R, et al. Deletion of the cold thermoreceptor TRPM8 increases heat loss and food intake leading to reduced body temperature and obesity in mice. *J Neurosci.* (2018) 38:3643–56. doi: 10.1523/JNEUROSCI.3002-17.2018
72. Millqvist E. TRPV1 and TRPM8 in treatment of chronic cough. *Pharmaceuticals* (2016) 9:45. doi: 10.3390/ph9030045
73. Lin MJ, Yang XR, Cao YN, Sham JS. Hydrogen peroxide-induced Ca²⁺ mobilization in pulmonary arterial smooth muscle cells. *Am J Physiol Lung Cell Mol Physiol.* (2007) 292:L1598–608. doi: 10.1152/ajplung.00323.2006
74. Crawford DC, Moulder KL, Gereau RWT, Story GM, Mennerick S. (2009). Comparative effects of heterologous TRPV1 and TRPM8 expression in rat hippocampal neurons. *PLoS ONE* 4:e8166. doi: 10.1371/journal.pone.0008166
75. Mergler S, Cheng Y, Skosyrski S, Garreis F, Pietrzak P, Kociok N, et al. Altered calcium regulation by thermosensitive transient receptor potential channels in etoposide-resistant WERI-Rb1 retinoblastoma cells. *Exp Eye Res.* (2012) 94:157–73. doi: 10.1016/j.exer.2011.12.002
76. Mergler S, Derckx R, Reinach PS, Garreis F, Bohm A, Schmelzer L, et al. Calcium regulation by temperature-sensitive transient receptor potential channels in human uveal melanoma cells. *Cell Signal* (2014) 26:56–69. doi: 10.1016/j.cellsig.2013.09.017
77. Pan R, Tian Y, Gao R, Li H, Zhao X, Barrett JE, et al. Central mechanisms of menthol-induced analgesia. *J Pharmacol Exp Ther.* (2012) 343:661–72. doi: 10.1124/jpet.112.196717
78. Alpizar YA, Boonen B, Gees M, Sanchez A, Nilius B, Voets T, et al. Allyl isothiocyanate sensitizes TRPV1 to heat stimulation. *Pflugers Arch.* (2014) 466:507–15. doi: 10.1007/s00424-013-1334-9
79. Ramachandran R, Hyun E, Zhao L, Lapointe TK, Chapman K, Hirota CL, et al. TRPM8 activation attenuates inflammatory responses in mouse models of colitis. *Proc Natl Acad Sci USA.* (2013) 110:7476–81. doi: 10.1073/pnas.1217431110

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Bräunig, Mergler, Jyrch, Hoefig, Rosowski, Mittag, Biebermann and Khajavi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.