

## Supplementary Material

### 1 SUPPLEMENTARY TABLES AND FIGURES

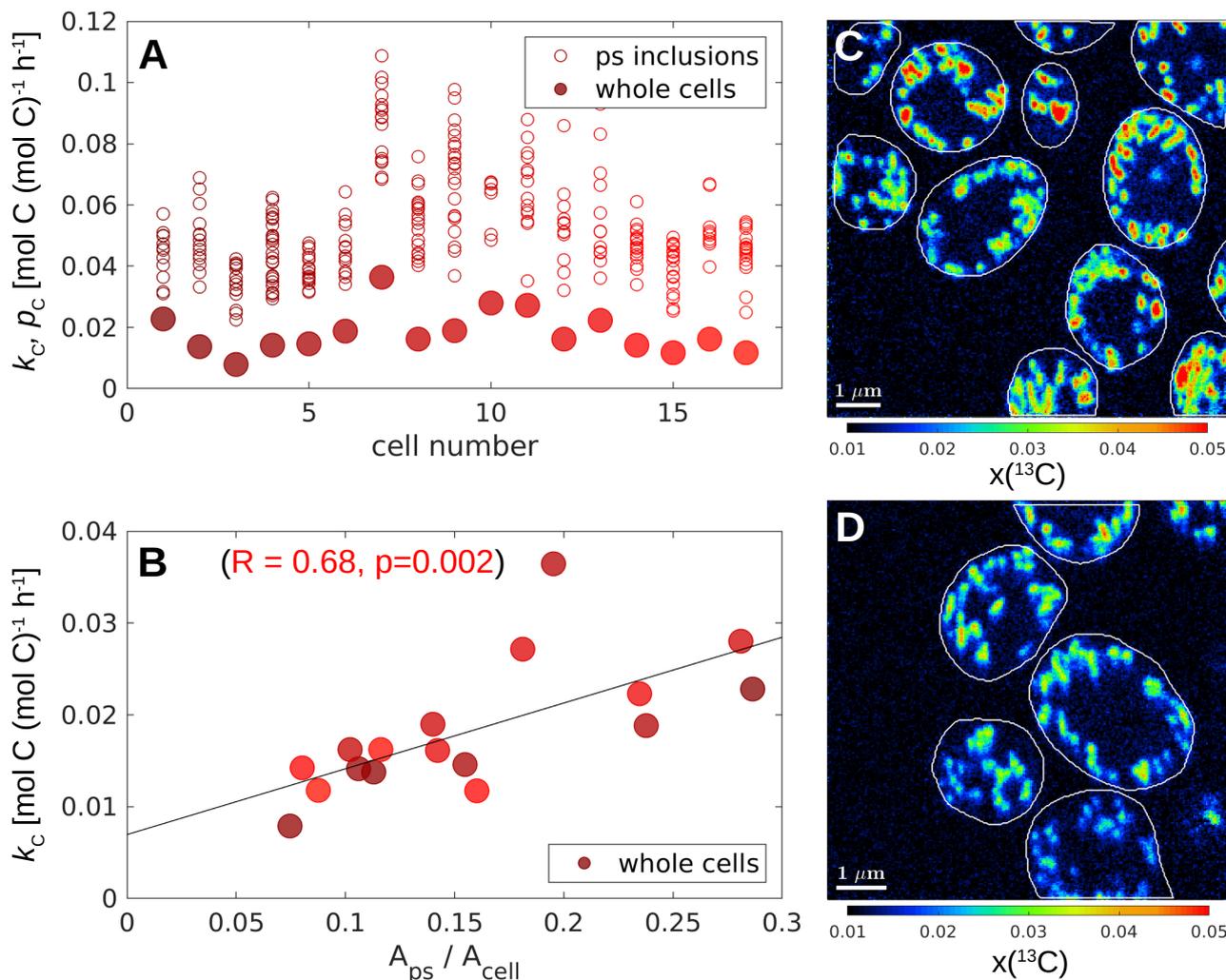
**Table S1.** Relevant chemical and isotopic characteristics of the incubation media used for SIP experiments.

Incubation period	Culture	DIC <sup>†</sup>		N <sub>2</sub> or NO <sub>3</sub>	
		[mmol L <sup>-1</sup> ]	$x(^{13}\text{C})_S$	[mmol L <sup>-1</sup> ]	$x(^{15}\text{N})_S$
07:30–09:30 (morning)	N <sub>2</sub>	1.4	0.23	0.3	0.06
	NO <sub>3</sub>	1.2	0.28	22.7	0.07
	SC-N <sub>2</sub>	1.4*	0.23	0.3	0.06
14:45–17:15 (afternoon)	N <sub>2</sub>	0.5	0.67	0.3	0.06
	NO <sub>3</sub>	0.4	0.86	22.7	0.07
	SC-N <sub>2</sub>	0.3	0.99	0.3	0.06
21:45–17:15 (night+day)	N <sub>2</sub>	0.6 <sup>§</sup>	0.53	0.3	0.06
	SC-N <sub>2</sub>	0.4 <sup>§</sup>	0.74	0.3	0.06
21:45–00:45 (early night)	N <sub>2</sub>	0.6 <sup>§</sup>	0.53	0.3	0.06
02:00–07:00 (late night)	N <sub>2</sub>	1.0 <sup>§</sup>	0.32	0.3	0.06
21:45–07:00 (full night)	N <sub>2</sub>	0.6 <sup>§</sup>	0.53	0.3	0.06
	NO <sub>3</sub>	0.5 <sup>§</sup>	0.68	16.2	0.10
	SC-N <sub>2</sub>	0.4 <sup>§</sup>	0.74	0.3	0.06

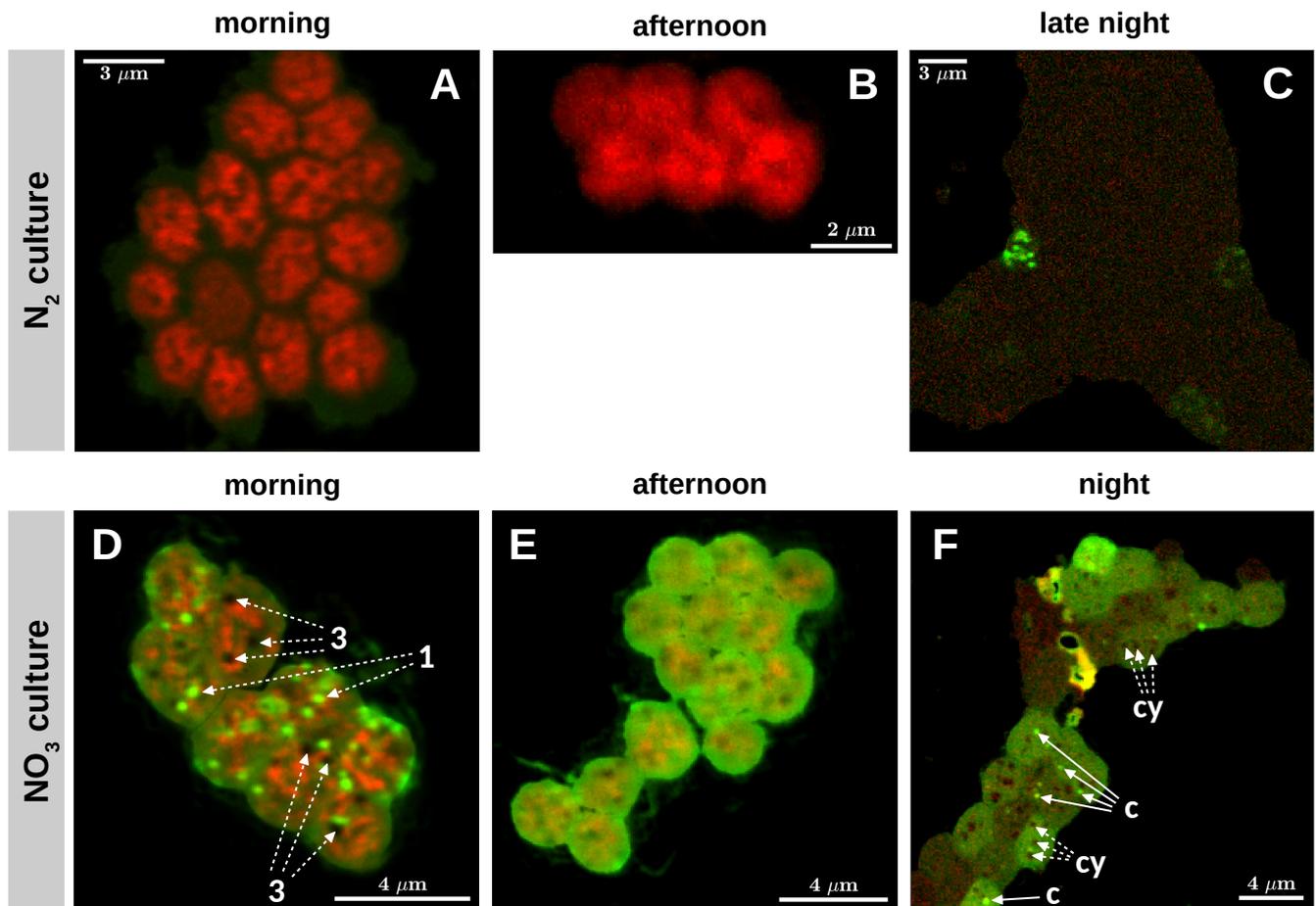
<sup>†</sup> DIC concentrations [mmol L<sup>-1</sup>] and <sup>13</sup>C atom fractions [ $x(^{13}\text{C})_S$ ] were calculated based on DIC measurements in samples from bioreactors taken at the same time point and the known amount of added NaH<sup>13</sup>CO<sub>3</sub>.

\* Value was assumed to be the same as in the N<sub>2</sub> culture.

<sup>§</sup> Values were interpolated from DIC concentrations measured at 07:00, 09:00 and 14:00.



**Supplementary Figure S1. Intra-cellular heterogeneity of C assimilation in N<sub>2</sub>-fixing *Cyanobacteria* 51142.** Shown are data for cells from the N<sub>2</sub> culture incubated in the morning (07:30–09:30). **(A)** Carbon-specific rates of C assimilation in individual polysaccharide (ps) inclusions ( $p_C$ ; open symbols) and in whole cells ( $k_C$ ; filled symbols), calculated for 279 inclusions in 17 selected cell sections.  $p_C$  varied among polysaccharide inclusions (CV = 32%), with 61% and 39% of the total variance explained by differences among and within cells, respectively ( $SS_{groups}/SS_{total} = 0.0455/0.0746 = 0.61$ ,  $SS_{error}/SS_{total} = 0.0291/0.0746 = 0.39$ ). Variation in  $p_C$  values among cells was significant (ANOVA,  $F(16, 262) = 25.6$ ,  $p = 10^{-44}$ ). **(B)** Correlation between  $k_C$  in whole cells and the relative area of the cell sections covered by polysaccharide inclusions,  $A_{ps}/A_{cell}$  (coefficient of determination  $R^2 = 0.46$ ). The correlation reveals that the cell-specific <sup>13</sup>C enrichment depends on the cell volume probed by nanoSIMS. **(C–D)** Examples of <sup>13</sup>C atom fraction images from which data in panels A–B were calculated. White lines show approximate cell outlines, <sup>13</sup>C-enriched spots correspond to polysaccharide inclusions.



**Supplementary Figure S2. Images of the isotopic composition of *Cyanothece 51142* cells.** Shown are additional overlays of the  $^{13}C$  (red) and  $^{15}N$  (green) atom fractions measured in cells from the  $N_2$  and  $NO_3$  culture deposited on polycarbonate membrane filters. Note that panel C shows cells from the  $N_2$  culture incubated during late night (02:00–07:00). In panels D and F, examples of cyanophycin inclusions (cy) and carboxysomes (c) are marked with dashed-line and solid-line arrows, respectively.